MAP3K1 function is essential for cytoarchitecture of the mouse organ of Corti and survival of auditory hair cells

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ABSTRACT

MAP3K1 is a serine/threonine kinase that is activated by a diverse set of stimuli and exerts its effect through various downstream effector molecules, including JNK, ERK1/2 and p38. In humans, mutant alleles of MAP3K1 are associated with 46,XY sex reversal. Until recently, the only phenotype observed in Map3k1tm1Yxia mutant mice was open eyelids at birth. Here, we report that homozygous Map3k1tm1Yxia mice have early-onset profound hearing loss accompanied by the progressive degeneration of cochlear outer hair cells. In the mouse inner ear, MAP3K1 has punctate localization at the apical surface of the supporting cells in close proximity to basal bodies. Although the cytoarchitecture, neuronal wiring and synaptic junctions in the organ of Corti are grossly preserved, Map3k1tm1Yxia mutant mice have supernumerary functional outer hair cells (OHCs) and Deiters’ cells. Loss of MAP3K1 function resulted in the downregulation of Fgf3, Fgf8, Fgf10 and Atf3 expression in the inner ear. Fgf3, Fgf8 and Fgf10 have a role in induction of the otic placode or in otic epithelium development in mice, and their functional deficits cause defects in cochlear morphogenesis and hearing loss. Our studies suggest that MAP3K1 has an essential role in the regulation of these key cochlear morphogenesis genes. Collectively, our data highlight the crucial role of MAP3K1 in the development and function of the mouse inner ear and hearing.

KEY WORDS: Map3k1, Mekk1, Fgf3, Fgf8, Fgf10, Hearing loss, Supernumerary outer hair cells, MAPK pathway, FGF signaling pathway

INTRODUCTION

Mitogen-activated protein kinases (MAPKs) are responsible for regulating a wide array of cellular functions and processes. The MAPK signaling cascade consists of three tiered phosphorylation steps, starting with the phosphorylation of MAPK kinase kinases (MAP3Ks, MEK kinases or MKKKs) in response to a plethora of stimuli, which in turn phosphorylate the MAPK kinases (MAP2Ks, MEK or MKKs) and then the MAPKs (Kyraki and Avruch, 2001; Uhlik et al., 2004). MAP3K1, a member of the MAPK kinase family, plays a diverse cell-signaling function in various biological systems, including immune system development and function (Gallagher et al., 2007; Labuda et al., 2006), vasculature remodeling (Li et al., 2005), tumor progression (Cuevas et al., 2006), cardiogenesis (Minamino et al., 2002), and injury repair (Deng et al., 2006).

MAP3K1 belongs to the serine/threonine kinase class that also participates in the regulation of the MAPK cascade (Cuevas et al., 2007; Hagemann and Blank, 2001; Uhlik et al., 2004). The MAP3K1 protein contains an ubiquitin interaction motif (UIM), a caspase-3 cleavage site and a conserved kinase domain (Uhlik et al., 2004; Witowsky and Johnson, 2003). MAP3K1 is associated with the plasma membrane and is tethered by α-actinin to actin stress fibers and by protein tyrosine kinase 2 (PTK2) to focal adhesions in cells (Christerson et al., 2002; Cuevas et al., 2003). Following an apoptotic signal, caspase-3 cleaves MAP3K1 at phosphorylated Asp874 (p.Asp874), resulting in the separation of the N-terminal UIM motif from the 91-kDa C-terminal kinase domain, releasing it from the cell membrane into the cytosol (Bonvin et al., 2002; Schlesinger et al., 2002). MAP3K1 is activated in response to a number of different stimuli, such as cold, growth factors, mild hyperosmolarity, microtubule disruption, cell shape disturbance, pro-inflammatory cytokines and other physiological stresses (Sadoshima et al., 2002; Xia et al., 2000; Yujiri et al., 1998). Once activated, MAP3K1 exerts its effect through the JNK, ERK1/2 and p38 MAPK pathways, as well as the transcription factors Jun and NF-κB (Bonvin et al., 2002; Guan et al., 1998; Xia et al., 2000; Yujiri et al., 1998).

In humans, mutant alleles of MAP3K1 are associated with 46,XY gonadal dysgenesis (Pearlman et al., 2010). These gain-of-function alleles affect the downstream phosphorylation of p38 and ERK1/2, as well as binding of MAP3K1 with the cofactors RHOA and MAP3K4 (Loke and Ostrer, 2012). Additionally, in vitro studies have suggested that these mutations alter the sex-determination pathway by concomitantly upregulating β-catenin expression and downregulating expression of the SOX9, SRY, FGF9 and FGF2 genes (Loke et al., 2014). However, mice carrying the Map3k1 loss-of-function allele display a minor testicular deficit in the developing gonad and have normal gross appearance besides the open-eyelid phenotype (Warr et al., 2011). MAP3K1-deficient mice display an open eye at birth (EOB) phenotype (Zhang et al., 2003) and have immune-system and wound-healing deficits, abnormal retinal vascularization, disintegration of retinal pigment epithelium, loss of photoreceptors, and retinal degeneration (Mongan et al., 2011). Additionally, cultured keratinocytes from these mutant mice display a lack of actin stress fiber formation and deficient cell migration (Yujiri et al., 2000; Zhang et al., 2003).

Previous studies have demonstrated the role of MAPK-mediated fibroblast growth factor (FGF) signaling in ontic induction and development (Urness et al., 2010). Hearing depends on the precise
**TRANSLATIONAL IMPACT**

Clinical issue

Hearing loss, which can present during early life or as a late-onset condition, is one of the most common neurosensory disorders worldwide. Normal hearing depends on the precise organization of sensory hair cells and non-sensory supporting cells within the organ of Corti (OC) in the cochlea of the ear. Any significant alterations to cell number, alignment or patterning within the OC causes hearing loss, underlining the importance of a defined cytoarchitecture in the OC for sound perception. Thus, understanding the molecular signaling cascades that lead to inner-ear sensory-cell differentiation and function is important for defining the molecular basis of hearing loss and devising strategies for hearing restoration. In this study, the putative role of MAP3K1 (a serine/threonine kinase with a pivotal function in MAPK signal transduction cascades) in inner-ear development and function was explored in mice.

Results

To determine whether MAP3K1 plays a part in hearing, the authors characterized homozygous Map3k1tm1Yxia kinase-null mutant mice. In the mouse inner ear, MAP3K1 is localized at the apical surface of cochlear supporting cells, in close proximity to basal bodies. A hearing test of the mutant mice revealed early-onset profound hearing loss, along with progressive degeneration of outer hair cells (OHCs). The authors show that the cytoarchitecture, neuronal wiring and synaptic junctions in the OC are unaffected; however, loss of MAP3K1 function results in an extra row of functional OHCs and Deiters’ cells (a type of cochlear supporting cell). Loss of MAP3K1 function also results in downregulation of members of the fibroblast growth factor (FGF) signaling pathway: Fgf3, Fgf8, Fgf10 and Atf3 expression in the inner ear. Previous studies have shown that Fgf3r, Fgf8 and Fgf10 have a role in induction of the otic placode – from which the auditory system develops during embryogenesis – or in otic epithelium development in mice.

Implications and future directions

Functional deficits in the FGF signaling pathway are known to cause defects in cochlear morphogenesis and hearing loss in mice. This study provides evidence that MAP3K1 has an essential role in the regulation of the FGF signaling pathway during the development, function and survival of inner-ear hair cells. Homozygous Map3k1tm1Yxia mice represent another model for the investigation of signaling pathways involved in hearing loss. In addition, elucidation of the central role of the MAP3K1 kinase protein could have implications for the controlled regeneration of inner-ear sensory cells for hearing restoration.

**RESULTS**

**MAP3K1 is localized with basal bodies in supporting cells**

To comprehend the role of Map3k1 in inner-ear development and function, we used previously generated Map3k1tm1Yxia mice (Xia et al., 2000; Zhang et al., 2003). In these mice, the exons encoding the kinase domain of MAP3K1 (Map3k1tm1Yxia) have an extra row of OHCs and supporting cells in the OC and suffer from early-onset profound hearing loss. Our results demonstrate the indispensable role of MAP3K1 in mouse inner-ear development and function. At the apical surface of supporting cells (Fig. 1B). Interestingly, the MAP3K1–β-galactosidase fusion protein and basal-body marker pericentrin colocalize with a very exquisite pattern (Fig. 1C). The MAP3K1–β-galactosidase fusion protein was expressed as two puncta on either side of the pericentrin-labeled basal cell bodies (Fig. 1C, inset). Basal bodies are known to direct planar cell polarity (PCP) of sensory hair cells (Jones et al., 2008). Therefore, we investigated the orientation of the OHC bundles in Map3k1 mutant mice, and no significant deficit was observed (see Fig. S1).

Additionally, weak diffused cytoplasmic expression of Map3k1 was also observed in the inner hair cells and outer hair cells (Fig. 1C), and in the marginal and intermediate cells of the stria vascularis (see Fig. S2A). Moreover, β-galactosidase staining in adult Map3k1 heterozygous mice revealed expression in supporting cells of the lesser epithelial ridge, greater epithelial ridge, stria vascularis, Reissner’s membrane and the spiral ganglion neurons (see Fig. S2B).

**Map3k1tm1Yxia/tm1Yxia mice have supernumerary outer hair cells and Deiters’ cells**

Upon confocal imaging of the OC from Map3k1tm1Yxia heterozygous and homozygous mice, we observed supernumerary outer hair cells (OHCs) throughout development (Fig. 2). The Map3k1tm1Yxia heterozygous mice have sparse one- to ten-cell stretches of an extra row of OHCs, whereas homozygous Map3k1tm1Yxia mice have a continuous extra row of OHCs in the apical, middle and basal cochlear turns (Fig. 2A). These supernumerary OHCs also have correctly polarized mechanosensitive hair bundles at their apical poles (Fig. 2B). Although no difference was observed in the number of inner hair cells (IHCs) (Fig. 2C), a statistically significant increase in OHCs was observed throughout the cochlear duct in homozygous and heterozygous Map3k1tm1Yxia mice (Fig. 2C). However, in comparison with homozygous mutant mice, fewer extra OHCs were observed throughout the cochlear duct in heterozygous Map3k1tm1Yxia mice (Fig. 2C), suggesting a dose-dependent role of MAP3K1-mediated signaling in the cytoarchitecture of the mouse OC.

In higher vertebrates, each hair cell in the OC is enveloped by supporting cells (Fig. 3A). In the mouse OC, the IHCs rest on the inner phalangeal cells, whereas each OHC rests upon a single Deiters’ cell, and inner and outer pillar cells separate these two types of sensory hair cells (Fig. 3A). To determine the effect of extra rows of OHCs on the precise cytoarchitecture of the OC, we immunolabeled the cochlear sections from postnatal day 0 (P0) control and Map3k1tm1Yxia mutant mice with an anti-Prox1 antibody, a marker for pillar and Deiters’ supporting cells. In Map3k1tm1Yxia mutant mice, the extra row of OHCs was also supported by an extra row of Deiters’ cells (Fig. 3B). However, the organization of the inner and outer pillar cells, Claudius cells and the tunnel of Corti was not affected in Map3k1tm1Yxia mutant mice (Fig. 3B, C). These findings suggest that MAP3K1 has a role in the development of controlled cytoarchitecture of mouse OC.

To assess the functional status of supernumerary hair cells, we briefly exposed (15 s) the OC explants from control and Map3k1tm1Yxia mutant mice to FM1-43, a steryl pyridinium dye that enters the hair cells via partially open mechanotransduction channels at rest (Gale et al., 2001; Meyers et al., 2003). Hair cells from control and Map3k1tm1Yxia mice showed dye loading in all rows of sensory cells, including the extra row of OHCs (Fig. 3D). We also performed synaptophysin immunostaining to identify postsynaptic endings of efferent neurons forming axodendritic organization of sensory hair cells and non-sensory supporting cells within the organ of Corti (OC). Any substantial alteration in the cellular number, alignment or patterning within the OC causes hearing loss, underlining the necessity of defined cytoarchitecture in the OC for sound perception. Here, we report that mice lacking the FGF signaling pathway during the development, function and survival of inner-ear hair cells. Homozygous Map3k1tm1Yxia mice represent another model for the investigation of signaling pathways involved in hearing loss. In addition, elucidation of the central role of the MAP3K1 kinase protein could have implications for the controlled regeneration of inner-ear sensory cells for hearing restoration.
synapses with the dendrites of spiral ganglion neurons in P12 control and mutant mice (Fig. 3E). Interestingly, the synaptophysin labeling was also observed at the base of an extra row of OHCs (Fig. 3E, arrowhead). Furthermore, no obvious difference in the neuronal wiring of the OC in the control and Map3k1tm1Yxia mutant mice was observed at P12 (Fig. 3F). However, the width of the area of neurons around the OHCs was wider in Map3k1tm1Yxia mutant mice, likely due to the wiring of the extra row of OHCs (Fig. 3F). Collectively, we observed that Map3k1tm1Yxia mutant mice have a completely developed, structurally supported, polarized and functional extra row of innervated OHCs and Deiters’ cells.

Map3k1tm1Yxia mutant mice have early-onset hearing loss

Next, to assess the hearing function of Map3k1tm1Yxia mutant mice, we measured auditory brainstem responses (ABRs) and distortion product otoacoustic emissions (DPOAEs). Although the wild-type control and Map3k1tm1Yxia heterozygous mice had comparable hearing thresholds across all the tested frequencies, significant hearing loss was observed in the homozygous Map3k1tm1Yxia mice at P16 and P30 (Fig. 4A,B). Similarly, the DPOAEs of wild-type and Map3k1tm1Yxia heterozygous mice were comparable, whereas the homozygous Map3k1tm1Yxia mouse had no detectable DPOAE thresholds (Fig. 4C). Taken together with ABRs, these results suggest that hearing loss in Map3k1tm1Yxia mutant mice is likely to result from peripheral (cochlear) deficiencies. In contrast, Map3k1tm1Yxia mutant mice did not exhibit any observable vestibular dysfunction phenotypes, such as hyperactivity, head-tossing or circling behavior.

Map3k1tm1Yxia mutant mice display rapid degeneration of OHCs and spiral ganglion neurons

Next, we examined the morphology of the cochlear epithelium at various developmental stages to determine the underlying cause of hearing loss observed in the Map3k1tm1Yxia mutant mice. Confocal imaging of myosin-VIIa-labeled OC of Map3k1tm1Yxia mice revealed normal development of IHCs and OHCs, which were indistinguishable from those of wild-type and heterozygous mice until P12 (Fig. 5). By P14, Map3k1tm1Yxia homozygous mice displayed obvious degeneration of OHCs in the basal turn, and by P16, no intact OHCs were observed in the basal turn (Fig. 5L). Furthermore, by P16, a varying degree of OHC degeneration was observed along the length of the cochlea (Fig. 5J-L). However, by P30, almost all of the OHCs in the basal and middle turn of the cochlea were degenerated, and many OHCs in the apical coil were also degenerated (Fig. 5M-O). In contrast, the IHCs appeared largely intact even at P75 (Fig. 5 and data not shown). The loss of
OHCs in Map3k1tm1Yxia mutant mice was followed by progressive degeneration of the spiral ganglion, which was more pronounced in the basal region at P90 (Fig. 6). Thus, the elevated hearing thresholds observed in MAP3K1-deficient mice are likely to be caused by the rapid degeneration of OHCs. These results suggest that MAP3K1 function is essential for the maintenance of OHCs in the mouse auditory system.

Genes of the FGF signaling pathway are downregulated in Map3k1 mutant mice

We reasoned that the extra row of OHCs and Deiters’ cells and degeneration of hair cells in Map3k1 mutant mice is likely to be caused by impaired intracellular signaling during the development and maturation of mouse OC. Therefore, we examined the expression of various genes associated with the Map3k1-mediated
Fig. 3. Cytoarchitecture of the organ of Corti (OC) is preserved in Map3k1tm1Yxia mutant mice. (A) Schematic representation of the developing mouse OC at P0 and P10. OHC, outer hair cell; IHC, inner hair cell; DC, Deiters’ cell; OPC, outer pillar cell; IPC, inner pillar cell; HC, Hensen’s cells; IPhC, inner phalangeal supporting cells. (B) Cross-section of Map3k1tm1Yxia mutant and wild-type control mice at P0, immunostained with Prox1 (green) and myosin VIIa (red). The arrows point to the pillar cells, whereas Deiters’ cells are marked by the arrowheads. Map3k1tm1Yxia mutant mice have an extra row of OHCs accompanied by an extra row of Deiters’ cells. Scale bar: 10 μm. (C) Immunostaining with the anti-CD44 antibody, a marker for OPCs, including Claudius cells, revealed an intact gross cytoarchitecture of the OC in Map3k1tm1Yxia mutant mice at P0 and P10. Scale bar: 10 μm. (D) No apparent difference in the FM1-43 dye uptake was observed among control and Map3k1tm1Yxia mutant explants. FM1-43 dye was also taken up by the supernumerary OHCs present in Map3k1tm1Yxia mutant mice. Scale bar: 10 μm. (E) Supernumerary OHCs in Map3k1tm1Yxia mutant mice are innervated and have synaptic junctions (arrowheads), immunolabeled with synaptophysin (green). Scale bar: 10 μm. (F) Map3k1tm1Yxia mutant mice have grossly intact neuronal wiring. Neurofilament (NF-200) protein immunostaining of wild-type control and Map3k1tm1Yxia mutant mice revealed grossly intact neuronal wiring of an extra row of OHCs. Scale bar: 10 μm.
signaling pathway (Fig. 7 and see Fig. S3) and transcriptional targets of FGF signaling, as well as the development-related genes in Map3k1tm1Yxia mutant mice at P10 (before the onset of hearing and OHC degeneration). As expected, the heterozygous mice had reduced expression of Map3k1, whereas the homozygous mice had no expression (Fig. 7). Among the 42 genes analyzed, we observed significant (P<0.001) downregulation of Aif3, Fgfr3, Fgf8 and Fgf10 in the inner ear of Map3k1 mutant mice (Fig. 7). Previous studies have shown that Fgfr3, Fgf8 and Fgf10 have a role in the otic placode induction or in otic epithelium development in mice, and their functional deficits cause defects in cochlear morphogenesis and hearing loss (Hayashi et al., 2007; Mansour et al., 2009; Pannier et al., 2009). Our results suggest that MAP3K1 has an essential role in the regulation of these key cochlear morphogenesis genes during development.

**DISCUSSION**

MAP3K1 is involved in the cellular response to a wide array of stimuli, including growth factors, pro-inflammatory cytokines, cell shape disturbance, microtubule disruption and a variety of cell stress signals (Cardone et al., 1997; Deak et al., 1998; Nakagami et al., 2001; Widmann et al., 1998; Xie et al., 2000; Yujiri et al., 1998), which translate into activation of the downstream JNK-, p38- and ERK-mediated MAPK pathways (Schlesinger et al., 1998; Xu et al., 1996). The full-length MAP3K1 protein is tethered to insoluble structures such as membranes or the cytoskeleton by the actin-bundling protein α-actinin (Christerson et al., 1999), and has also been found to be associated with actin fibers entering focal adhesions, so is implicated in controlling their turnover (Cuevas et al., 2003). MAP3K1 interacts directly with the signaling molecule MEK1 (Karandikar et al., 2000), which, in the mouse oocyte, has been associated with the control of microtubule organization and colocalizes with the centrosomal protein γ-tubulin (Yu et al., 2007). Interestingly, in Map3k1tm1Yxia mice, the MAP3K1–β-galactosidase fusion protein in the supporting cells of the OC also colocalized with the centrosomal protein and, therefore, might also participate in microtubule organization in the inner ear non-sensory cells and in PCP (Jones et al., 2008). There are many examples of PCP proteins, such as Vangl2, that are not expressed in the inner ear sensory hair cells, but still severely affect the stereocilia bundle orientation (Montcouquiol et al., 2006). However, in the Map3k1tm1Yxia mutant mice we did not observe any significant stereocilia bundle orientation deficit, which might reflect that Map3k1 has either no direct role in PCP, or that there is functional redundancy with other family members.

In a parallel study, a splice-site mutation in the Map3k1 gene in goya mutant mice was identified as part of an ENU mutagenesis screening program (Parker et al., 2015). goya mice also exhibit a progressive hearing-loss phenotype along with the supernumerary OHCs. Interestingly, when maintained on the same genetic background (C3H), the goya and Map3k1tm1Yxia mutant mice were found to have a similar progressive hearing-loss phenotype and were profoundly deaf by 9 weeks (Parker et al., 2015). Modulation of the phenotype of a given allele by the genetic background of an incompletely penetrant trait is a well-documented phenomenon (Doetschman, 2009; Montagutelli, 2000). Intriguingly, in addition to the increased hearing loss that we observed, a study involving Map3k1 mice on the C57BL/6J background has also shown a drastic decrease in the number of animals surviving to maturity (Warr et al., 2011), which further supports the notion of a Map3k1 modifier gene.
Although the expression of MAP3K1 was observed in the supporting cells of the OC, Map3k1tm1Yxia mutant mice exhibited four rows of OHCs. During development, extra OHCs can be found in wild-type mice, but they are generally restricted in the apical turns of the cochlea, and their frequency can vary depending on different inbred mouse strains (Lim and Anniko, 1985), whereas, in Map3k1tm1Yxia mutant mice, the fourth row of OHCs was found throughout development and along the length of the cochlea. One possibility to explain such a phenotype includes the lack of apoptosis of a population of cells in the prosensory domain that would otherwise have undergone degeneration during development. Normally, full-length MAP3K1 acts as an anti-apoptotic protein but, when cleaved by caspase 3, becomes pro-apoptotic (Schlesinger et al., 2002). Relocalization of the MAP3K1 C-terminal 91-kDa fragment containing the kinase domain is necessary for its pro-apoptotic function; however, in the case of Map3k1tm1Yxia mice, the kinase domain is replaced with the lacZ domain, rendering it incapable of inducing apoptosis in the supporting cells of the developing OC.

Alternatively, the supernumerary OHCs in Map3k1tm1Yxia could stem from the deficit in the control of the prosensory domain size during embryogenesis. The sensory epithelial cell differentiation initiates with specification of the prosensory domain in the otocyst. Many signaling molecules, including the sonic hedgehog, Notch and Wnt pathway genes, act in concert to form a highly specialized patterned sensory epithelium. Canonical Wnt (Wnt/β-catenin) signaling is also essential for the specification of the otic placode because either conditional deletion or activation of β-catenin in Pax2-positive ectodermal cells or Foxg1-positive placodal cells results in a substantial reduction or expansion of the size of the otic placode, respectively (Freyer and Morrow, 2010; Ohyama et al., 2006). MAP3K1, via direct interaction with Axin1, is known to...
modulate Wnt/β-catenin signaling pathway activity (Sue Ng et al., 2010). Previously, fibroblasts from Map3k1tm1Yxia mutant mice displayed a tenfold increase in transcription factor Lef/Tcf reporter activity in response to Wnt3a expression compared to an only threefold increase observed in wild-type cells (Jin et al., 2013). Because Wnt expression regulates proliferation in the early prosensory domain and hair cell differentiation in the later stage, the level of Wnt/β-catenin signaling can affect hair cell formation in a dose-dependent manner (Jacques et al., 2012). It is plausible that, during development, Wnt/β-catenin signaling might have resulted in relative overactivation of downstream transcription factors, leading to the expansion of the prosensory domain and excessive differentiation of OHCs in Map3k1tm1Yxia mutant mice. Future work using cell-specific markers and cochlear tissue from various embryonic developmental stages will provide knowledge regarding the role of MAP3K1 in sensory-cell specification and differentiation.

Map3k1tm1Yxia mutant mice exhibit downregulation of at least four genes, which are known for their role in inner-ear development and function. ATF3, a transcription factor that is induced in response to a number of stress stimuli (Hai et al., 1999; Kyriakis et al., 1994; Liang et al., 1996), is significantly downregulated in Map3k1tm1Yxia mutant mice. ATF3 is also implicated in the survival, repair and neurite outgrowth in association with heat shock protein 27, Akt and Jun (Nakagomi et al., 2003; Pearson et al., 2003). Also, Fgfr3 mutant cochlea show neuronal wiring pattern disruption (Puligilla et al., 2007). However, in Map3k1tm1Yxia mutant mice, we did not observe any gross deficit in the neuronal wiring. Furthermore, the DPOAE data are suggestive of a functional deficit of hair cells in Map3k1tm1Yxia mutant mice. These results suggest that downregulation of Atf3 is unlikely to be the reason for the supernumerary OHCs and Deiters’ cells, and hearing deficits, observed in Map3k1tm1Yxia mutant mice.

Before the onset of hair-cell degeneration at P10, we found significant downregulation of Fgfr3, Fgf8 and Fgf10 expression in the Map3k1tm1Yxia mutant mouse OC cells. Dysfunction of Fgfr3 in mice, either due to loss (Hayashi et al., 2007) or gain (Mansour et al., 2009; Pannier et al., 2009) of function, results in hearing loss. Interestingly, both Fgfr3 mutant alleles have supernumerary OHCs (Hayashi et al., 2007; Mansour et al., 2009), as is observed in Map3k1tm1Yxia mutant mice. This implies that, in inner-ear development, Map3k1 and Fgfr3 might participate in the same signaling cascade to control the precise cytoarchitecture of the OC. However, in contrast to Fgfr3 mutant mice, which have either loss of pillar or Deiters’ supporting cells in the OC (Hayashi et al., 2007; Mansour et al., 2009; Pannier et al., 2009), Map3k1tm1Yxia mutant mice
mice do not exhibit any obvious deficit in the differentiation of pillar cells; rather, they have an extra row of Deiters’ cells. The phenotypes of overproduction of OHCs and loss of pillar cells reported for the gain-of-function allele of Fgfr3 phenotype is remarkably similar to Sprouty2, an antagonist of FGF signaling, mutant mice (Shim et al., 2005). Intriguingly, the hearing-loss phenotype in Sprouty2 mutant mice can be partially rescued by genetically reducing the Fgf8 expression level (Shim et al., 2005). Furthermore, reduction of Fgf10 expression reverted the fate-switched supporting cells back and restored hearing in Fgfr3 mutant mice (Mansour et al., 2013). Besides Fgfr3, Fgfr8 and Fgf10 are also downregulated in Map3k1tm1Yxia mutant mice, which could account for the grossly intact supporting cells in these mutant mice as compared to Fgfr3 mutant mice.

Our studies further highlight the complexity of the signaling pathway(s) required for formation of the precise cytoarchitecture of mouse OC and the maintenance of OHCs. We also found that MAP3K1 function was necessary for regulation of the FGF-mediated pathway in the auditory system and hearing in mice.

MATERIALS AND METHODS

Map3k1 mutant mice

Map3k1tm1Yxia mice were generated as described previously (Xia et al., 2000; Zhang et al., 2003). In summary, the targeting vector replaced the entire kinase domain in the Map3k1 locus, resulting in the formation of a MAP3K1–β-galactosidase fusion protein. These targeted embryonic stem (ES) cells (Map3k1tm1Yxia) were injected into mouse blastocysts, and the resulting chimeras were crossed with C57BL/6 mice to obtain mice with germ-line transmission of the Map3k1tm1Yxia allele. All experiments were approved by the Animal Care and Use Committees at the University of Maryland, School of Medicine in accordance with the National Institutes of Health (NIH) Guide for the Care and Use of Laboratory Animals.

ABR and DPOAE measurements

Hearing function was evaluated by ABR analyses at two different time points (P16 and P30) in mice of all the three genotypes, i.e. wild-type, heterozygous and Map3k1tm1Yxia homozygous mice. Mice were anesthetized with intraperitoneal injections of Avertin (0.4-0.75 mg/kg body weight, Sigma-Aldrich, St Louis, MO). All recordings were performed in a sound-attenuated chamber using an auditory-evoked potential diagnostic system RZ6 (Tucker-Davis Technologies Inc., Alachua, FL) as previously described (Nayak et al., 2013). Experiments represent the mean and standard error of mean (s.e.m.) of three or more animals. Significance was analyzed using Student’s t-test.

DPOAE recordings were performed at P30 with an acoustic probe (ER-10C, Etymotic Research, Elk Grove Village, IL) using a DP2000 DPOAE measurement system version 3.0 (Starkey Laboratory, Eden Prairie, MN). Two primary tones, with a frequency ratio of f2/f1=1.2, where f1 represents the first tone and f2 represents the second, were presented at intensity levels L1=65 dB sound pressure level (SPL) and L2=55 dB SPL. Also, f2 was varied in one-eighth octave steps from 8 to 16 kHz. DP grams comprised 2f1–f2 DPOAE amplitudes as a function of f2.

Confocal imaging

The temporal bones from the control and mutant mice were isolated and fixed for 1 h at room temperature or overnight at 4°C in 4% paraformaldehyde (Electron Microscopy Sciences, Hatfield, PA), followed by three washes with phosphate buffered saline (PBS). Finely dissected cochlear coils were permeabilized and blocked in 5% BSA, 2% normal goat serum and 0.1% Triton X-100 in PBS for 1 h. The tissue samples were washed and probed with primary antibody overnight at 4°C. We used the following primary antibodies in our studies: β-galactosidase (1:1000 dilution; MP Biomedicals, Solon, OH), myosin VIIa (1:200; Proteus BioSciences, Ramona, CA), synaptophysin (1:200; Abcam, Cambridge, MA), NF-200 (1:200; Sigma-Aldrich), Prox1 (1:100; Millipore, Billerica, MA) and Pericentrin (1:200; Millipore, Billerica, MA). After three washes with PBS, samples were probed with a fluorescently labeled Alexa-Fluor-488 or -546 secondary antibody (1:500; Life Technologies, Grand Island, NY) for 1 h at room temperature. Rhodamine-phalloidin or Alexa-Fluor-647-conjugated phalloidin (1:250; Life Technologies, Grand Island, NY) was used to label actin. Samples were mounted using ProLongGold (Life Technologies, Grand Island, NY) and viewed under an LSM 700 confocal microscope (Zeiss Microimaging Inc., Thornwood, NY) using a ×63, 1.4 N.A. oil-immersion lens.

Scanning electron microscopy (SEM)

For SEM studies, the inner ears were isolated at P14 and fixed for 1.5 h in a fixative containing 2.5% glutaraldehyde and 2 mM CaCl2 in 0.1 M sodium cacodylate buffer, and later washed three times with 0.1 M sodium cacodylate buffer. The inner ears were post-fixed in 1% osmium tetroxide in 0.1 M sodium cacodylate buffer for 1 h at room temperature. After washing three times with PBS buffer, inner ears were decalcified by incubating in 0.25 M EDTA for 1-2 days at 4°C. The samples were then finely dissected to expose the sensory epithelium and to remove the tectorial membrane and the stria vascularis. The cochlear tissues were then dehydrated in gradient alcohol changes, critical point dried, sputter coated with platinum, and imaged on a field-emission SEM.

Cochlear explants and FM1-43 dye uptake

OC explants from P3 wild-type and Map3k1tm1Yxia mutant mice were finely dissected and cultured on collagen-coated glass-bottom Petri dishes (MatTek Corporation, Ashland, MA) in DMEM medium supplemented with 10% fetal bovine serum (Life Technologies, Grand Island, NY) and 10 mg/ml ampicillin (Millipore, Billerica, MA) at 37°C and 5% CO2. The explants were kept in vitro for 2 days to allow for complete adhesion to the dish. To test the FM1-43 dye (N-[3-triethylammoniumpropyl]-4-[4- (dibutylamino)-styryl]pyridinium dibromide) (Life Technologies, Grand Island, NY), uptake by the hair cells, the culture was exposed to 3.0 μM FM1-43 in Hank’s Balanced Salt solution (HBSS) for 15 s and then quickly washed three times with HBSS. The culture was mounted with Fluoro-Gel (Electron Microscopy Sciences, Crofton, MD) and immediately imaged under an LSM 700 confocal microscope (Zeiss Microimaging Inc.).

Cryosections

Hemisectioned heads of P0 mice or inner ears of P10 and older mice were collected and fixed in 4% paraformaldehyde (PFA) at 4°C overnight. For older mice, the temporal bones were decalcified in 0.25 M EDTA for 1 to 2 days at 4°C. The inner ears were then equilibrated with 30% sucrose in PBS overnight at 4°C, embedded in OCT, and immediately frozen by placing the block on an ethanol-dry-ice mix. The frozen tissue blocks were sectioned with a cryostat at 14 μm thickness.

Reverse transcriptase PCR (RT-PCR) and real-time PCR

Total RNA was isolated from finely dissected OC from P10 inner ear tissue of wild-type, heterozygous and Map3k1tm1Yxia homozygous mice (five mice each) using the RiboPure RNA isolation kit (Life Technologies, Grand Island, NY) and cDNA was prepared using an oligo-dT primer and SMARTScribe Reverse Transcription enzymes (Clontech, Mountain View, CA). To determine the differential expression of various genes, SYBR-Green-based real-time primers (available upon request) were designed using Integrated DNA Technologies online PrimerTime qPCR assay design tool (http://www.idtdna.com/Scitools/Applications/RealTimePCR/). The real-time PCR assays were performed in triplicate using an ABI StepOnePlus Real-Time thermal cycler (Life Technologies, Grand Island, NY). Ct values were normalized using Gapdh and actin as an endogenous control, and fold changes of different genes were calculated using SABiosciences online software (http://pcedataanalysis.sabiosciences.com/pcr/arrayanalysis.php). The genes with a twofold change and with a P-value less than 0.05 based on a Student’s t-test analysis were considered significant.

X-gal staining

The inner ears were isolated from wild-type and Map3k1tm1Yxia heterozygous mice at P30 and were fixed for 10 min at room temperature
in LacZ fixative (1% formaldehyde, 0.2% glutaraldehyde and 0.02% NP-40 in PBS). Then, the samples were washed twice with PBS containing 0.02% NP-40 and 2 mM MgCl₂, 10 min each, followed by incubation at 37°C overnight in a staining solution [1 mg/ml X-Gal, 5 mM K₃Fe(CN)₆, 5 mM K₄Fe(CN)₆ and 0.1 M MgCl₂]. Following staining, samples were washed in PBS, decalcified and cryosectioned as described above. The sections were counterstained with hematoxylin and eosin and imaged using a ×40 oil immersion lens on a Zeiss Axioplan Apotome-equipped microscope.

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Competing interests
The authors declare no competing or financial interests.

Author contributions
R.Y., R.B.H., Z.M.A. and S.R. designed the research. R.Y., Y.X., Z.M.A. and S.R. and/or contributed new reagents and performed research. R.Y., Q.M., R.B.H., C.P., Y.X., Z.M.A. and S.R. analyzed data. All authors drafted or revised the manuscript, and all authors approved the final version.

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Supplementary information
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