The challenge of cholangiocarcinoma: dissecting the molecular mechanisms of an insidious cancer

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Cholangiocarcinoma is a fatal cancer of the biliary epithelium and has an incidence that is increasing worldwide. Survival beyond a year of diagnosis is less than 5%, and therapeutic options are few. Known risk factors include biliary diseases such as primary sclerosing cholangitis and parasitic infestation of the biliary tree, but most cases are not associated with any of these underlying diseases. Numerous in vitro and in vivo models, as well as novel analytical techniques for human samples, are helping to delineate the many pathways implicated in this disease, albeit at a frustratingly slow pace. As yet, however, none of these studies has been translated into improved patient outcome and, overall, the pathophysiology of cholangiocarcinoma is still poorly understood. There remains an urgent need for new approaches and models to improve management of this insidious and devastating disease. In this review, we take a bedside-to-bench approach to discussing cholangiocarcinoma and outline research opportunities for the future in this field.

Cholangiocarcinoma: the clinical problem

Epidemiology

Cholangiocarcinoma (CCA) is the second most common primary liver cancer globally, and the most common cause of death from primary liver cancer in the United Kingdom, where it kills 1500 people annually (Khan et al., 2012). At diagnosis, 65% of cases of CCA are in individuals who are over 65 years old, and the disease occurs equally in men and women. Globally, CCA incidence varies, reflecting differences in genetic and environmental risk factors. The highest incidence is in north-east Thailand (with 80-90 cases per 100,000 people), whereas Australia has the lowest incidence (with 0.4 cases per 100,000 people). Notably, the incidence of CCA is increasing worldwide for unknown reasons (Khan et al., 2012).

Classification

CCA is a primary malignancy, usually adenocarcinomatous, arising in the bile ducts that carry bile from its site of production in the liver to its site of action in the small intestine (Fig. 1). Topographically, CCA is classified as intrahepatic (IHCC), arising in the liver, or extrahepatic (EHCC), arising outside the liver. IHCC is subcategorised morphologically into mass-forming, periductular-infiltrating or intraductal, the latter being the least common but with a more favourable prognosis; however, IHCC tumours can possess a combination of these characteristics (Lim and Park, 2004). EHCC is subcategorised into perihilar CCA, involving the bifurcation of the main ducts (50% of all CCA), and distal EHCC. IHCC, distal EHCC and perihilar CCA are increasingly regarded as each having a distinct epidemiology, pathogenesis and management requirements. However, clarifying these distinctions has been hampered by inconsistent classification, especially the inclusion of perihilar lesions as either IHCC or EHCC (Khan et al., 2012) and sometimes including gallbladder carcinoma or hepatocellular carcinoma in these classifications. Where known, relevance to subtypes will be made clear in this review.

Risk factors for CCA

In the Western world, 80% of CCA cases are sporadic and have no identifiable risk factor (Khan et al., 2012). Smoking, alcohol consumption, diabetes and obesity have not been consistently shown to increase risk, although a small contribution cannot be ruled out (Tyson and El-Serag, 2011). Risk factors that have been identified are generally associated with chronic biliary inflammation. For example, primary sclerosing cholangitis (PSC; see Box 1 for Glossary) is associated with 10% of CCA cases in the Western world (La Russo et al., 2006). In patients with PSC, the risk of developing CCA is 1% per annum over 10 years, with a cumulative lifetime risk of 9-31% – 1500-fold that of the general population (Burak et al., 2004; Claessen et al., 2009). In the management of this high-risk group, because of the often desmoplastic nature of CCA it is challenging to discriminate between malignancy and the benign strictures characteristic of PSC. Liver cirrhosis of mixed aetiology conveys a tenfold relative risk (Sorensen et al., 1998). Uncommon abnormalities of biliary anatomy, such as choledochal (bile duct) cysts, intrahepatic biliary cysts (Carolii’s syndrome), biliary papillomatosis and adenoma, are associated with a high lifetime risk of CCA of 6-30% (Tyson and El-Serag, 2011). This high rate of malignant transformation warrants prophylactic resection (Tyson and El-Serag, 2011). Congenital or acquired abnormalities of pancreatic biliary-duct junctions allow pancreatic reflux and resultant chronic cholangitis, increasing risk of CCA.
**Clinical Puzzle**

and Clonorchis sinensis biliary infestation with liver flukes (including associated with CCA. In areas of south-east (Tyson and El-Serag, 2011). This is difficult to advise either biliary resection (for isolated cholangiocarcinoma) or liver transplant (for it, are proving difficult for him to adjust to. Without a more conclusive test for cholangiocarcinoma, it is unlikely that additional cases associated with this agent will emerge (Tyson and El-Serag, 2011). Several different infections are also associated with CCA. In areas of south-east Asia, where the incidence of CCA is high, biliary infestation with liver flukes (including Opisthorchis viverrini, Clonorchis sinensis and Schistosomiasis japonica) is endemic. High faecal egg loads and high titres of fluke-specific antibodies are associated with a 14- and 27-fold increase in CCA risk, respectively, although the mechanism of cholangiocarcinogenesis is unknown (Hughes et al., 2006). The prevalence of hepatitis C (HCV) and/or hepatitis B (HBV) is several times higher in the CCA population than the general population (Kobayashi et al., 2000; Donato et al., 2001; Chuang et al., 2009; Zhou et al., 2008; Lee et al., 2009a), and infection with HCV conveys a hazard ratio of 2.55 for IHCC (El-Serag et al., 2009). Cryptosporidiosis, a common human pathogen, has been implicated in both CCA and gastric cancer (Certad et al., 2012). Finally, chronic carriers of typhoid have a sixfold increase in CCA incidence (Abdel Wahab et al., 2007).

**Presentation and diagnosis**

CCA usually presents late in the course of disease, with general malaise and symptoms of biliary obstruction such as jaundice and biliary sepsis. Thereafter, the patient usually deteriorates quickly and, without treatment, death tends to occur from sequelae of obstruction. Currently, diagnosis is based on a combination of modalities, but is still sometimes only confirmed by a resection specimen. The only currently available serum biomarker, carbohydrate antigen 19-9 (Ca19-9; also known as sialylated Lewis antigen), is a poor diagnostic marker, with sensitivity of 40-70%, specificity of 50-80% and positive predictive value of 16-40% (reviewed by Khan et al., 2012). This is in part because 10% of individuals lack the Lewis antigen so cannot produce Ca19-9. Moreover, Ca19-9 can be expressed in any obstructive jaundice, and cannot therefore discriminate CCA from pancreatic or gastric malignancy. Imaging (ultrasound, MRI, MRCP, CT) can detect biliary obstruction, stricture and mass lesions, but cannot confirm malignancy. Cytology from endoscopic retrograde cholangiopancreatography (ERCP) brushings fails to demonstrate malignancy in 50% of cases, owing to the desmoplastic nature of CCA (Khan et al., 2005); this can be improved to 40-70% by combining with targeted biopsies using cholangioscopy in specialist centres (Khan et al., 2012). In PSC strictures, the occurrence of polysomy [detected by fluorescent in-situ hybridisation (FISH)] is 88% specific for CCA (Bangaruvelgam et al., 2010), and can precede radiological and pathological changes by up to 2.7 years (Barr Fritcher et al., 2011). However, FISH is not yet part of routine workup in all centres, including those in the UK.

**Treatment**

Treatment options for CCA are few, and include surgery, limited pharmacotherapeutics and endobiliary therapies. Surgical resection is the only curative treatment for CCA (Friman, 2011), but a minority of cases are amenable to surgery, with most precluded by metastasis. Post-resection 5-year survival is only 20-30% (reviewed by Rosen et al., 2010). Liver transplantation was previously associated with poor survival and is hence contraindicated. However, recent data from the United States have shown that transplantation with neoadjuvant chemoradiotherapy in a carefully selected patient subgroup results in a 70% 5-year survival, and is therefore a re-emerging treatment option (Darwish Murad et al., 2012).

Because more than 50% of patients have lymph node metastases at presentation, effective chemotherapeutic regimes are vital in treating CCA. The UK NCRI ABC-01 and ABC-02 (Phase III) trials demonstrated that combination chemotherapy with cisplatin and gemcitabine improved progression-free survival and quality of life (Valle et al., 2009), although overall survival remained poor. The efficacy of adjuvant chemotherapy after resection is unproven, although ongoing trials will report soon (e.g. BILCAP, http://public.ukcrn.org.uk, UKCRN ID 1473).

In unresectable CCA of the main or common ducts, endoscopic stenting allows drainage, which relieves itch and sepsis. However, tumour ingrowth and obstruction frequently occur, and repeat procedures are...
common if the patient survives beyond 6 months. Improvements in stent technology could decrease procedure frequency. Endobiliary radio-frequency ablation (RFA) coupled with stenting has been shown to be safe (Steel et al., 2011), and the results of long-term efficacy studies are eagerly awaited.

To improve the outlook for individuals with CCA, both clinical and bench science are imperative. In the rest of this article, we first review existing models of CCA, and then go on to discuss the complex and interrelated aspects of CCA pathophysiology.

**Currently available models**

**Cell lines**

Cholangiocarcinoma cell lines were first reported in 1985, when HChol-Y1 (immortalised from a patient with IHCC) was characterised and was shown to be xenotransplantable and to increase serum Ca19-9 and CEA (carcinoembryonic antigen, a marker raised in several cancers including pancreatic, hepatic and gastrointestinal) in recipient mice (Yamaguchi et al., 1985). More recently, the non-tumorigenic rat cholangiocyte line BDE1 has been transformed to constitutively express activated Neu, the rat homologue of the receptor tyrosine kinase ErbB2 (see below). Although results from tissue studies of ErbB2 expression in human tumours are inconsistent, elevated ErbB2 is reportedly present in bile from patients suffering from CCA (Su et al., 2001), and in the ducts of high-risk patients (i.e. those with PSC, heptolithiasis, etc.) (Endo et al., 2002). ErbB2 promotes carcinogenesis by activating p42/44 mitogen-activated protein kinase (MAPK) signalling (driving CCA cell proliferation), as well as phosphatidylinositol 3-kinase (PI3K)-Akt signalling (increasing CCA cell survival and resistance to apoptosis) (reviewed by Sirica, 2008). The BDEneu line is highly malignant and shows the molecular features of moderately differentiated CCA (Lai et al., 2005). It is also highly amenable to in vitro studies. The BDE1 parent line has also given rise to a less aggressive phenotype, by selection of spontaneous transformants developing after repeated cycling (Sirica et al., 2008). The availability of two malignant lines derived from one parent line offers unique opportunities for comparative studies.

Although several other CCA cell lines have been developed (see Table 1), standard cell culture methods do not reflect the molecular milieu, microenvironment and architecture inhabited by the native cholangiocyte. Thus, researchers are turning to more sophisticated methods, including tissue-engineering and 3D modelling, to study CCA. In EHCC research, bile-duct tissue engineering has focussed on producing a functional prosthetic conduit for bile flow, rather than on modelling pathology (Miyazawa et al., 2012). In IHCC, the Sirica group recently reported a novel organotypic model using a 3D collagen gel matrix to support co-cultured clonal CCA cells and clonal α-smooth muscle actin (α-SMA)-positive stromal cancer-associated fibroblasts (CAFs). The cells used to form this model were derived from orthotopic IHCC formed by bile-duct inoculation of tumorigenic rat BDEsp transformed cholangiocytes (Campbell et al., 2012; Sirica et al., 2009). α-SMA-positive human stromal cell lines have also been extracted from patient resection specimens (Utispan et al., 2010), and establishing a similar organotypic system with human-derived tumour and stromal cells is a worthwhile aim.

**Animal models**

Although animal models of CCA exist, none recapitulate the human disease. In particular, few models carrying CCA-related mutations have been explored, with the exceptions being p53-deficient mice, which readily develop CCA on exposure to carbon tetrachloride (Farazi et al., 2006), and mice with liver-specific deficiency of SMAD4 and PTEN, which develop IHCC (Xu et al., 2006). Other genes known to be commonly mutated in human CCA, such as KRAS, have yet to be fully explored in modelling work (Hezel et al., 2010). Animal models of PSC are similarly elusive, as recently reviewed (Pollheimer et al., 2011).

In terms of modelling IHCC, xenografts created by transplanting human or cell-line material into rodents have been used extensively in assessing potential therapeutic agents; unfortunately, the results in such models correlate poorly with clinical outcome (reviewed by Sausville and Burger, 2006; Voskoglou-Nomikos et al., 2003; Fava, 2010). Earlier models of CCA relied on natural cholangiocarcinogenesis of inoculated *Opisthorchis viverrini* to induce biliary malignancy in rodents (Tesana et al., 2000), or the carcinogenic compounds furan, thioacetamide and dimethylnitrosamine (Lai and Sirica, 1999; Yeh et al., 2004; Thamavit et al., 1993). More recently, biliary inoculation of rodents with tumorigenic BDEneu or BDEsp cells has been shown to result in IHCC growth, reflecting progressive and early disease, respectively (Sirica et al., 2008). Although application of sorafenib in
these parallel models demonstrated therapeutic benefit and allowed mechanistic explorations (Blechacz and Gores, 2008), a recent Phase II trial of sorafenib in humans failed to demonstrate benefit and was terminated early (El-Khoueiry et al., 2012). This trial in particular has reinforced the limited translatability of current models. The generation of new models that enable delineation of important signalling pathways in CCA, and others that emulate important CCA risk factors (such as PSC), will provide a more reliable reflection of CCA pathology and allow us to better predict the effects of new interventions.

**Human samples**

Despite the fact that working with human samples has ‘real world’ relevance, there is large inter-patient variability, and controlling for ‘real world’ variables is an ongoing challenge. Nevertheless, human samples are particularly relevant for researching biomarkers. There have been several recent advances in this area by analysing various types of samples and using several techniques. Metabolomic analysis of bile by nuclear magnetic resonance (NMR) has demonstrated CCA-specific expression profiles, with alterations in genes including various HOX genes, collagens, *PVT1*, *MUC4*, *MUC5AC* and *LEF1* (Chapman et al., 2012). Other recent advances include improvements in proteomic analysis of bile fluid by LC-MS/MS (Farina et al., 2011; Zabron et al., 2011) and capillary electrophoresis (Lankisch et al., 2011). Finally, NMR-based small-molecule analysis of urine identified diagnostic candidates for hepatocellular carcinoma (Shariff et al., 2011); a similar approach could be successful in CCA. These findings might form the basis of future biomarker panels, although further work is still required before they can be translated to the clinic.

**Current understanding of CCA pathophysiology**

Dissection of the underlying molecular mechanisms of CCA has begun to reveal that a background of chronic inflammation – providing a high rate of cell turnover and a rich milieu of cytokines and growth factors – allows the accumulation of mutations (Table 2) and the proliferation of mutated cells. CCA can also arise in the absence of chronic inflammation, however, through mutations that alter pathways in a manner similar to chronic inflammation. These alterations include stimulation of angiogenesis, as well as changes in various mechanisms (e.g. cell migration, interactions with stromal tissues, exposure to bile acids, cell cycle and apoptosis). Some of these inflammatory and non-inflammatory mechanisms are described below.

**Angiogenesis: VEGF**

CCA requires a rich vascular bed. Although minimal information is available on angiogenesis in CCA, it is known that high levels of vascular endothelial growth factor (VEGF) are expressed by various human CCA cell lines (KMC-1, KMC-2, KMBc and KMG-C) and tumour tissues (Ogasawara et al., 2001). Studies of xenografted cells have shown that endothelin 1 (ET-1) inhibits
VEGF-A and VEGF-C expression or release, which reduces cell proliferation and increases fibrosis and apoptosis in tumour tissue. The ET-1 receptor antagonist bosentan has been suggested as a possible therapeutic agent (Fava et al., 2009). It has been shown that increased VEGF might operate via an autocrine loop to increase malignant cell proliferation (Gaudio et al., 2006) in response to increased histamine production by the activation of HHR receptors by histidine decarboxylase (HDC). It has been suggested that interrupting this autocrine loop by inhibiting HDC might offer therapeutic benefit (Francis et al., 2012).

**EMT in CCA**

Epithelial-mesenchymal transition (EMT) is a key process in the development of many cancers, resulting in cellular rearrangements and a motile fibroblastic phenotype that is adapted to invasion. EMT has recently been demonstrated in IHCC, and mirrors that seen in other malignancies, with cells having an aggressive, dedifferentiated phenotype and increased motility. Epithelial markers E-cadherin and α- and β-catenin are downregulated in these cells, whereas markers such as N-cadherin, S100A4 and vimentin, are upregulated (Sirica et al., 2009; Yao et al., 2012; Seol et al., 2011). These changes correlate with increased invasiveness in vitro, and with metastasis and poorer prognosis in vivo (Sirica et al., 2009; Fabris et al., 2011; Yao et al., 2012).

Both epidermal growth factor (EGF) (see below) and transforming growth factor β (TGFβ) signalling are implicated in EMT in CCA. In other cancers, receptor binding by TGFβ causes SMAD2 and SMAD3 phosphorylation and their heterodimerisation with SMAD4. These heterodimers translocate to the nucleus and regulate target gene transcription, resulting in upregulated expression of Snail, Slug and Twist, which in turn promote EMT (reviewed by Bierie and Moses, 2006). In CCA cell lines, TGFβ alters cell morphology from epithelial to mesenchymal, induces Snail and vimentin expression (Sato et al., 2010), and triggers a ‘cadherin-switch’ from E- to predominantly N-cadherin expression (Araki et al., 2011). When CCA cell lines are implanted as xenografts, TGFβ treatment increases invasiveness (Sato et al., 2010). In addition, the change to N-cadherin predominance in tissue correlates with disease progression and poor prognosis (Araki et al., 2011).

The receptor tyrosine kinase Met and its ligand, hepatocyte growth factor (HGF; also known as scatter factor), are also associated with EMT. Both of these proteins are more highly expressed in CCA than in normal tissue, and Met expression is associated with increased cell migration and invasiveness (Furge et al., 2000; Lee et al., 2009b). Met binding to HGF triggers a network of intracellular signals that accelerate tumour growth and spread (reviewed by Socoteanu et al., 2008).

**MMPs in CCA**

In addition to EMT, several other processes might promote cell motility and invasiveness in CCA. In particular, CCA is associated with increased levels of matrix metalloproteinases (MMPs), which break down the extracellular matrix (ECM) to allow tumour spread. Immunohistochemical analysis found that 48% and 76% of surgically resected specimens expressed MMP-9 and MMP-7, respectively (Itatsu et al., 2009; Itatsu et al., 2008), with MMP-7 being expressed by the malignant cholangiocytes themselves. Plasma MMP-7

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**Table 1. Cell lines derived from human and animal CCA**

<table>
<thead>
<tr>
<th>Cell line</th>
<th>Derived from</th>
<th>Construction</th>
<th>Comments</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>BDEsp</td>
<td>Rat cholangiocyte BDE1</td>
<td>Spontaneous transformation</td>
<td>Slow-growing; models early disease</td>
<td>Sirica et al., 2008b</td>
</tr>
<tr>
<td>BDEneu</td>
<td>Rat cholangiocyte BDE1 containing rat Neu oncogene</td>
<td>Retroviral vector Glu664-neu</td>
<td>Rapid-growing; models advanced disease</td>
<td>Lai et al., 2005</td>
</tr>
<tr>
<td>Choi-CK</td>
<td>Human IHCC</td>
<td>Cultured</td>
<td>Well-differentiated CCA</td>
<td>Kim et al., 2001</td>
</tr>
<tr>
<td>HCho1-Y1</td>
<td>Human resection specimen</td>
<td>Cultured</td>
<td></td>
<td>Yamaguchi et al., 1985</td>
</tr>
<tr>
<td>HuCC-T1</td>
<td>Human cholangiociarcinoma cells from malignant ascites</td>
<td>Cultured</td>
<td>Secrete Ca19-9, carbohydrate antigen 125, carcinoembryonic antigen, thrombin plasminogen activator</td>
<td>Miyagiwa et al., 1989</td>
</tr>
<tr>
<td>HuH-28</td>
<td>Human resected CCA</td>
<td>Cultured</td>
<td>Slow growing</td>
<td>Kusaka et al., 1988</td>
</tr>
<tr>
<td>ICBD-1</td>
<td>Human EHCC</td>
<td>Cultured</td>
<td>Poorly differentiated adenocarcinoma</td>
<td>Takiyama et al., 1998</td>
</tr>
<tr>
<td>KKU-100</td>
<td>Opisthochiasis-associated human EHCC</td>
<td>Cultured</td>
<td>Poorly differentiated tubular adenocarcinoma</td>
<td>Sripa et al., 2005</td>
</tr>
<tr>
<td>KMBC</td>
<td>Human EHCC</td>
<td>Serial transplant to nude mouse; cultured</td>
<td></td>
<td>Yano et al., 1992</td>
</tr>
<tr>
<td>SCK</td>
<td>Human IHCC</td>
<td>Cultured</td>
<td>Sarcomatous</td>
<td>Kim et al., 2001</td>
</tr>
<tr>
<td>SK-ChA-1</td>
<td>Human EHCC malignant ascites</td>
<td>Cultured</td>
<td></td>
<td>Knuth et al., 1985</td>
</tr>
<tr>
<td>SNU-245</td>
<td>Human EHCC</td>
<td>Cultured</td>
<td></td>
<td>Ku et al., 2002</td>
</tr>
<tr>
<td>SNU-1196</td>
<td>Human invasive hilar EHCC</td>
<td>Cultured</td>
<td>Carries deletions in p15,p16,p53</td>
<td>Sripa et al., 2005</td>
</tr>
<tr>
<td>SNU-1079</td>
<td>Human IHCC</td>
<td>Cultured</td>
<td>Carries deletions in p15,p16,</td>
<td>Saijyo et al., 1995</td>
</tr>
<tr>
<td>TFK-1</td>
<td>Human EHCC, papillary adenocarcinoma and tubular adenocarcinoma</td>
<td>Cultured</td>
<td></td>
<td>Watanabe et al., 2000</td>
</tr>
<tr>
<td>TK</td>
<td>Human cholangio carcinoma ascites</td>
<td>Cultured</td>
<td></td>
<td>Mao et al., 1990</td>
</tr>
<tr>
<td>WB-F344</td>
<td>Rat epithelial stem-like cells</td>
<td>Chemical and spontaneous transformation</td>
<td>Gives rise to a variety of CCA subtypes and other malignancies on transplant</td>
<td>Lai et al., 2005</td>
</tr>
</tbody>
</table>

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was found to be increased in obstructed perihilar and intrahepatic CCA, and was suggested as a putative diagnostic biomarker with AUC 0.73 (Leeawat et al., 2009). Elevated tumour MMP-7 has also been suggested to be a marker of poor post-operative prognosis (Itatsu et al., 2008). Notably, MMP-9 requires stabilisation by binding to neutrophil gelatinase-associated lipocalin (NGAL), which is reportedly increased in bile in patients suffering from malignant pancreaticobiliary obstruction (Zabron et al., 2011). NGAL knockdown in RMCCA1 cells (a CCA cell line) showed decreased invasiveness and migration in vitro (Nuntagowat et al., 2010).

### EGFR and ErbB2

The ErbB receptor kinase family [of which epidermal growth factor receptor (EGFR) is the founding member] are potent and broad-spectrum mediators of cholangiocarcinogenesis. They interact with multiple other factors that contribute to cholangiocarcinogenesis, including COX-2, interleukin-6 (IL-6), VEGF and Met, activating not only their own signalling pathway but enhancing that of others. Although this makes understanding the part played by individual factors more challenging, it also suggests that manipulation of ErbB activity with novel therapies could have a wide-reaching therapeutic benefit.

The four members of the ErbB family each have an extracellular ligand-binding domain, a transmembrane domain and an intracellular kinase domain. Ligand binding induces homo- or heterodimerisation, leading to transphosphorylation of the cytoplasmic tail, which regulates downstream signalling molecules. EGFR (ErbB1) binds several soluble ligands, including EGF, transforming growth factor-α (TGFα) and amphiregulin. In CCA, EGF operates through EGFR to activate p42/44 MAPK, extracellular regulated kinase (ERK) and PI3K-Akt pathways (thereby stimulating cell survival and inhibit apoptosis (reviewed by Sirica, 2008). As found for EGFR, ErbB2 homologue Neu in the rat cell line ERB2 homologous to Neu in the rat cell line demonstrates increased expression of ErbB2 and PI3K-Akt pathways (thereby stimulating cellular proliferation), as well as to promote cell survival and inhibit apoptosis (reviewed by Sirica, 2008). As found for EGFR, ErbB2 homologue Neu in the rat cell line shows increased ErbB2 expression in CCA tissue and bile fluid (Endo et al., 2001; Su et al., 2001). Expression of ErbB2 is also increased in tissues of patients with PSC and hepatolithiasis (risk factors for CCA) (Endo et al., 2002).

### Stromal factors

One of the characteristic histological findings in CCA is a dense desmoplastic stroma surrounding malignant glandular structures. This stroma is mostly made up of CAFs, which are histologically identified by immunoreactivity for α-SMA. In IHCC, increased numbers of α-SMA-positive CAFs

### Table 2. Selected genetic alterations associated with CCA for different cellular processes

<table>
<thead>
<tr>
<th>Gene</th>
<th>Incidence</th>
<th>Functional outcome</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Proliferation</strong></td>
<td></td>
<td></td>
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</tr>
<tr>
<td>KRAS</td>
<td>50% of IHCC cases, 30% of EHCC cases, 30% of PSC cases</td>
<td>Gain-of-function mutations cause activation of the RAS pathway and increased proliferation</td>
<td>Ahrendt et al., 2000</td>
</tr>
<tr>
<td>BRAF</td>
<td>22% of CCA cases</td>
<td>Hot-spot substitution mutations cause activation of the MAPK pathway, increasing proliferation</td>
<td>Tannapfel et al., 2003</td>
</tr>
<tr>
<td>KEAP1</td>
<td>11% of gallbladder CCA cases</td>
<td>Mutations cause aberrant activation of NF-E2-related factor 2, and increased expression of phosphorylated p38-MAPK increases cell proliferation</td>
<td>Shibata et al., 2008</td>
</tr>
<tr>
<td>RASSF1A</td>
<td>65-69% of CCA cases</td>
<td>Promoter methylation causes activation of the RAS pathway, increasing proliferation and dysregulating cell-cycle control</td>
<td>Wong et al., 2002</td>
</tr>
<tr>
<td>EGFR</td>
<td>13-15% of CCA cases</td>
<td>Activating substitution and deletion mutations increase proliferation via multiple pathways (see main text for details)</td>
<td>Leone et al., 2006</td>
</tr>
<tr>
<td><strong>Invasion and metastasis</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>CDH1</td>
<td>Promoter mutations in 11% of IHCC cases</td>
<td>Promoter mutations are associated with invasion and metastasis</td>
<td>Endo et al., 2001</td>
</tr>
<tr>
<td>AXIN1</td>
<td>Substitution mutations in 41% of IHCC cases</td>
<td>Mutations increase invasion and metastasis</td>
<td>Tokumoto et al., 2005</td>
</tr>
<tr>
<td><strong>Cell-cycle dysregulation</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>SMAD4</td>
<td>45% of IHCC cases</td>
<td>Substitution mutations cause cell-cycle dysregulation through the TGFβ pathway</td>
<td>Kang et al., 2002</td>
</tr>
<tr>
<td>CDKN2A (p16INK4A)</td>
<td>17-86% CCA cases</td>
<td>Loss-of-function mutation or methylation causes TP53 inactivation and loss of cell-cycle control</td>
<td>Tannapfel et al., 2000a</td>
</tr>
<tr>
<td>TP53</td>
<td>37% IHCC cases</td>
<td>Dysregulation of cell-cycle arrest, resistance to apoptosis</td>
<td>Tannapfel et al., 2000b</td>
</tr>
<tr>
<td><strong>Chronic inflammation</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>SOCS3</td>
<td>60% of IHCC cases</td>
<td>Silencing by promoter methylation removes suppression of IL-6–STAT3 activation</td>
<td>Isomoto et al., 2007</td>
</tr>
</tbody>
</table>
correlate with larger tumours and shorter patient survival, and conditioned medium from α-SMA-expressing CAFs stimulates greater proliferation of CCA cell lines than conditioned medium from other stromal cell populations (Campbell et al., 2012). Recently, it has been recognised that the stroma can directly influence the progression of CCA. A detailed review on this topic has recently been published (Sirica, 2012), so we limit our discussion below to the key points.

CAFs from CCA are known to produce various factors that can promote tumour progression. For example, perioxidin promotes the survival and invasion of CCA cell lines in vitro, and expression of perioxidin in resected tumours is a poor prognostic factor (Utispan et al., 2010; Sirica et al., 2009). Tenascin expression is also correlated with poor IHCC prognosis after resection, although its mechanism in IHCC is not clear (Sirica et al., 2009). Thrombospondin-1 overexpression in CAFs correlates with hypo-vascularity and metastasis (Tang et al., 2006), whereas galectin-1 expression is associated with dedifferentiation (Shimomishi et al., 2001). More is known about the mechanism of stromal-cell-derived factor-1, which is overexpressed at the CCA invasion front. Here, it activates the MEK1/2 and Akt pathways in IHCC cells via its cognate receptor C-X-C chemokine receptor 4 (CXCR4) (Leelawat et al., 2007). In addition, it has been shown that the mechanism whereby CAF-derived HGF promotes CCA cell motility and invasion in vitro involves E-cadherin dislocation and activation of the PI3K-Akt pathway (Menakongka and Suthiphonchai, 2010). WNT1-inducible-signalling pathway protein 1 (WISP1) is overexpressed in 49% of IHCC cases, and in situ hybridisation has shown WISP1 mRNA expression in CAFs rather than in the malignant cells (Tanaka et al., 2003). This overexpression correlated with lymphatic and perineural invasion, and with poor prognosis and survival. Moreover, WISP1-conditioned medium was shown to stimulate migration of HuCCT1 CCA cells in vitro (Tanaka et al., 2003). It has been suggested that targeting these stromal factors in combination with CCA cell factors could improve therapeutic efficacy (Sirica, 2012).

**Bile acids**

Bile acids, which are produced by hepatocytes from cholesterol, are transported to the small intestine in the bile, via the biliary tree, constantly bathing the biliary endothelium. In obstruction and cholestasis, which are associated with several risk factors for CCA, bile acids accumulate abnormally. Although bile acids have long been known to assist in nutrient absorption, their active roles in the regulation (and pathology) of the biliary endothelium in health and disease are the topic of recent and ongoing research. In particular, the bile-acid component deoxycholic acid activates EGFR (via TGFα) and induces COX-2 expression, activating p42/44 and p38 MAPK, as well as c-jun-N-terminal kinase (JNK) (Yoon et al., 2004); as discussed, activation of these pathways contributes to CCA. Deoxycholic acid also prolongs the activity of myeloid-cell leukaemia-1 (Mcl-1; a member of the pro-survival Bcl-2 protein family), promotes cellular proliferation and conveys resistance to Fas-induced apoptosis (Werneburg et al., 2003). However, not all bile acids are carcinogenic; for example, taurosodeoxycholate (TUDCA) inhibits human CCA growth in vitro (Alpini et al., 2004). Further understanding of the mechanisms by which bile acids contribute to CCA might reveal targets for therapy.

**Apoptosis and cell survival**

Suppression of the normal mechanisms that eliminate cells that accumulate DNA damage is a crucial step in carcinogenesis. In CCA, this suppression generally involves mutation or dysregulation of proteins such as p53 and the anti-apoptotic protein Bcl-2. In CCA cell lines, Bcl-2 is overexpressed (Tadlock and Patel, 2001), preventing caspase-3 activation and conferring resistance to apoptosis. p53 mutation is seen in 20-61% of CCA cases (Khan et al., 2003; Nault and Zucman-Rossi, 2011), dysregulating cell-cycle arrest, allowing Bcl-2 expression and suppressing the normal apoptotic response. Tumour necrosis-factor-related apoptosis-inducing ligand (TRAIL)-mediated apoptosis is reduced in CCA via mechanisms involving Notch-1 and cyclooxygenase-2 (COX-2) (see below) (Kobayashi et al., 2005). TGFβ regulates cell proliferation positively or negatively, mediated by down- or upregulation of the cyclin-dependent kinase inhibitor p21 (also known as Waf1), respectively (Miyazaki et al., 1998). Promoter methylation, which reduces transcription, and loss-of-function mutations in the cell-cycle regulators p14ARF and p16INK4a, and in β-catenin, occur less frequently in CCA than in other cancers, but when present are thought to promote cell survival (Tannapfel et al., 2002).

**Mechanisms associated with inflammation**

An inflammatory milieu contributes to the development of CCA. High concentrations of inflammatory mediators cause or allow the accumulation of mutations in proto-oncogenes, DNA mismatch-repair genes and tumour suppressor genes, and directly promote cell proliferation. In particular, expression of activation-induced cytokine deaminase (AID), a member of the DNA-RNA editing family, is induced by pro-inflammatory cytokines via the nuclear factor-κB (NFκB) pathway, and AID is present at high levels in PSC and CCA biopsies. Aberrant AID expression in biliary cells results in somatic mutations in p53, MYC and p16INK4a, potentiating malignant transformation (Komori et al., 2008). There is also evidence that cell-mediated immune surveillance might be deficient in CCA. Analysis of single nucleotide polymorphisms (SNPs) in CCA in patients with underlying PSC identified two polymorphisms in NKG2D (natural killer group 2 D receptor), which is expressed in NK cells and T lymphocytes: carriers of either of these SNPs have odds ratios above 2 for developing CCA (Melum et al., 2008). Such polymorphisms remain to be identified in sporadic or fluke-related CCA.

In the sections below, we review the role of some specific inflammatory mediators and pathways in CCA.

**IL-6**

The cytokine IL-6 is dramatically upregulated in cultured CCA cells compared with normal biliary tract cells, and is regarded as a growth and survival factor (Sugawara et al., 1998; Isomoto et al., 2005). IL-6 has multiple effects in promoting CCA (Fig. 2). In normal cholangiocytes, activation of the Janus kinase-signal transducer and activator of transcription (JAK-STAT) pathway by IL-6 enhances transcription of suppressor of cytokine signalling 3 (SOCS3), which mediates a classic negative feedback loop for IL-6 production. In CCA, SOCS3 expression is decreased (Isomoto et al., 2007). Methylation of the SOCS3 promoter at CpG islands is increased in tumour tissues and in human CCA cell lines (Mz-ChA-1 and CCLP1), which correlates with decreased transcription-factor binding and SOCS3
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Fig. 2. IL-6 contributes to CCA through multiple pathways, linking chronic inflammation with the development of malignancy. IL-6-mediated activation of STAT-3 confers resistance to apoptosis via upregulating Mcl-1. SOCS3, a negative regulator of IL-6 signalling, seems to be downregulated in CCA. IL-6-mediated upregulation of p38 MAPK leads to dysregulation of the cell cycle, whereas increased PGRN increases cellular proliferation. IL-6 also alters the expression and activity of various miRNAs (only some are shown here), at least in part by upregulating DNMT1 and increasing miRNA methylation. These changes lead to downregulation of Rassf1a and p16INK4a, and derepression of MAP3K8, causing the dysregulation of cell-cycle control.

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transcription (Isomoto et al., 2007). *In vitro*, demethylation re-establishes SOCS3 expression and decreases IL-6-dependent STAT3 phosphorylation (Isomoto et al., 2007). IL-6 is known to induce DNA methyltransferase-1 (DNMT1) expression in CCA cell lines via miRNAs (see below), and DNMT1 causes epigenetic modification of several oncogenes (Braconi et al., 2010). It has been demonstrated in colon cancer arising in ulcerative colitis (another inflammation-associated malignancy) that SOCS3 is silenced by DNMT1 (Li et al., 2012); it would be interesting to investigate whether this is also the case in CCA.

IL-6-induced phosphorylation and activation of STAT3 causes transcriptional upregulation of Mcl-1 (Kobayashi et al., 2005), which desensitises CCA cells to TRAIL-mediated apoptosis, an effect that is reversed when autonomous IL-6 signalling is interrupted by SOCS3 re-expression (Isomoto et al., 2007). Disrupting this pathway is a promising therapeutic strategy: indeed, a multikinase inhibitor, sorafenib, which induces STAT3 dephosphorylation, resensitised CCA cells to TRAIL-mediated apoptosis both *in vitro* and in an orthotopic model (Blechacz et al., 2011). However, this agent did not have the same effect in early human trials (El-Khoueiry et al., 2012).

IL-6 induces increased expression of progranulin (PGRN) in CCA compared with non-malignant cholangiocytes (Frampton et al., 2012). Increased PGRN expression is seen in several tumour types, and correlates with high tumorigenicity and in some cases poor prognosis (e.g. epithelial ovarian cancer (Han et al., 2011)). In CCA cell lines, increased PGRN expression is independent of the JAK-STAT pathway, and instead occurs in response to IL-6-mediated phosphorylation of ERK1 or ERK2 and subsequent activation of ribosomal S6 kinase-1 (RSK-1). Activated RSK-1 phosphorylates the transcription factor C/ERBP, which binds the PGRN promoter (Frampton et al., 2012). Furthermore, recombinant PGRN increased proliferation of the same CCA cell lines through an Akt-dependent pathway. Notably, Akt is also involved in several pathways promoting CCA tumorigenesis that are independent of PGRN. For example, prostaglandin E2 (derived from COX-2 activation during inflammation) promotes CCA cell growth and invasion through prostaglandin E2 receptor EP1-mediated activation of Akt and EGF (Han and Wu, 2005). Furthermore, liver-specific disruption of the tumour suppressors SMAD4 (also known as DPC4) and PTEN leads to increased Akt phosphorylation and the development of CCA (Xu et al., 2006).

In addition to its anti-apoptotic and pro-proliferative roles, increased IL-6 production in CCA also dysregulates cell-cycle control through activating p38 MAPK, which decreases expression of the cell-cycle regulator p21 and is mitogenic (Tadlock and Patel, 2001).

**NO and COX-2**

Several lines of evidence support a role for COX-2 in cholangiocarcinogenesis. COX-2 is a common inflammatory mediator, whose expression is induced by cytokines and lipopolysaccharide, resulting in the synthesis of inflammatory prostaglandins. COX-2 directly links cholestasis and inflammation, because oxysterols (derived from cholesterol and present at high levels in cholestatic bile) stabilise COX-2 mRNA in human cholangiocytes (Yoon et al., 2004). Cholangiocytes in PSC, where risk of developing CCA is high, have high levels of COX-2 compared with another chronic inflammatory biliary disease, primary biliary cirrhosis (PBC), where CCA risk is normal (Sirica, 2005). Overexpression of COX-2 or treatment with prostaglandin E2 (a downstream mediator of COX-2) enhances the growth of human CCA cells, an effect that is reversed by COX-2 antisense depletion (Han et al., 2004). In CCA cell lines, oxysterol-induced COX-2 accumulation results in cellular proliferation and inhibition of apoptosis, whereas COX-2 inhibitors (e.g. celecoxib) induce apoptosis and decrease Akt phosphorylation (Yoon et al., 2004; Zhang et al., 2004). Celecoxib also inhibits proliferation in CCA cells via activation of the cyclin-dependent kinase inhibitors p21 and p27 (KIP1), arresting cells at G1-S (Han et al., 2004).

In *in vitro* experiments with a mouse CCA cell line indicate that cholangiocyte COX-2 expression is regulated in part via inducible nitric oxide synthase (iNOS) and nitric oxide (NO) (Ishimura et al., 2004). iNOS is expressed in response to inflammatory cytokines. Indeed, *Opisthorchis viverrini* expresses iNOS, although whether this promotes CCA remains to be seen.
Clinical and basic research opportunities

- To develop animal models with mutations in key genes implicated in cholangiocarcinogenesis.
- To apply new developments in materials science and nanotechnology to develop better in vitro 3D multilayered models of IHC and EHCC.
- To explore the relationship between the liver fluke and its endobiliary habitat, and identify factors that trigger malignant transformation in the host.
- To develop in vivo and in vitro models of PSC that reflect both the benign inflammatory condition and its contribution malignant transformation.
- To combine bioinformatics with multiple biological, biochemical and biophysical techniques to analyse human samples of bile, blood and protein and develop accurate diagnostic tests.
- To apply expanding knowledge of cholangiocarcinogenesis to drug development for CCA.

The future

Recent years have seen an expansion in basic research on CCA, meaning that pathways of cholangiocarcinogenesis are now beginning to be delineated. However, we are still unable to provide an early and definitive diagnosis of CCA, to predict which patients with high-risk conditions will go on to develop the disease, or to provide effective targeted therapy for any patient group. Thus, more work is urgently needed to identify diagnostic biomarkers and to design effective pharmacotherapies using existing and new models. A multidisciplinary approach that brings together biologists and engineers, clinicians and statisticians, parasitologists and materials scientists, among others, is required to tackle this insidious disease.

In analysing human samples, multi-modal analysis of multiple sample types, together with application of systems biology and bioinformatics approaches, is most likely to identify the next generation of biomarkers and diagnostic tests. Although various in vivo and in vitro models have been developed, they have thus far fallen short in translating useful ideas to the clinic. New animal models carrying specific mutations will provide important insights, but humanised in vitro models that mirror the molecular ecosystem of cancer-stroma interactions in a 3D matrix will probably provide the most significant advances. In terms of treatment strategies, our rapidly expanding understanding of genetic mechanisms will hopefully be translated into therapeutic siRNA- or miRNA-based drugs, possibly allowing a local application and avoiding systemic side effects.

The insidious and complex nature of CCA will continue to challenge us in understanding its pathophysiology, and the need for new models, innovative approaches and talented researchers is immense.

COMPETING INTERESTS

The authors declare that they do not have any competing or financial interests.

AUTHOR CONTRIBUTIONS

A.Z. drafted the original manuscript with help from R.J.E. S.A.K. provided the structure and overall management of the manuscript.

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