A mouse model for inherited renal fibrosis associated with endoplasmic reticulum stress

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ABSTRACT
Renal fibrosis is a common feature of renal failure resulting from multiple etiologies, including diabetic nephropathy, hypertension and inherited renal disorders. However, the mechanisms of renal fibrosis are incompletely understood and we therefore explored these by establishing a mouse model for a renal tubular disorder, referred to as autosomal dominant tubulointerstitial kidney disease (ADTKD) due to missense uromodulin (UMOD) mutations (ADTKD-UMOD). ADTKD-UMOD, which is associated with retention of mutant uromodulin in the endoplasmic reticulum (ER) of renal thick ascending limb cells, is characterized by hyperuricemia, interstitial fibrosis, inflammation and renal failure, and we used targeted homologous recombination to generate a knock-in mouse model with an ADTKD-causing missense cysteine to arginine uromodulin mutation (C125R). Heterozygous and homozygous mutant mice developed reduced uric acid excretion, renal fibrosis, immune cell infiltration and progressive renal failure, with decreased maturation and excretion of uromodulin, due to its retention in the ER. The ER stress marker 78 kDa glucose-regulated protein (GRP78) was elevated in cells expressing mutant uromodulin in heterozygous and homozygous mutant mice, and this was accompanied, both in vivo and ex vivo, by upregulation of two unfolded protein response pathways in primary thick ascending limb cells from homozygous mutant mice. However, this did not lead to an increase in apoptosis in vivo. Thus, we have developed a novel mouse model for renal fibrosis, which will be a valuable resource to decipher the mechanisms linking uromodulin mutations with ER stress and renal fibrosis.

KEY WORDS: Kidney, Fibrosis, ER stress, Mouse model, Endoplasmic reticulum

INTRODUCTION
Renal fibrosis is an integral factor in progression to end-stage renal failure (ESRF), regardless of the primary etiology, which may include diabetic nephropathy, hypertension or monogenic disorders (Eddy, 2014). However, mechanisms of renal fibrosis are incompletely understood, and there are currently no therapies to reverse or slow down the progression of renal fibrosis (Eddy, 2014). Autosomal dominant tubulointerstitial kidney disease (ADTKD), a monogenic cause of renal fibrosis, is characterized by urine concentrating defects, interstitial nephropathy with immune cell infiltration and glomerulosclerosis, and progressive loss of renal function leading to ESRF (Eckardt et al., 2015). Mutations in four genes have been identified to cause ADTKD, with the most common being in the UMOD gene, encoding uromodulin (Eckardt et al., 2015; Hart et al., 2002; Turner et al., 2003), referred to as ADTKD-UMOD. ADTKD-UMOD was previously referred to as familial juvenile hyperuricemic nephropathy type 1 (FJHN1; OMIM #162000), medullary cystic kidney disease type 2 (MCKD2; OMIM #603860) and glomerulocystic kidney disease (GCKD; OMIM #609886) (Eckardt et al., 2015; Rampoldi et al., 2003). In addition to the general clinical characteristics of ADTKD, patients with ADTKD-UMOD present with elevated serum urate concentrations due to low fractional excretion of uric acid (FEUA).

Uromodulin, a 640 amino acid glycosylphosphatidylinositol (GPI) anchored protein, is only expressed in the thick ascending limb (TAL) of the loop of Henle (Rampoldi et al., 2011). Uromodulin is synthesized within the endoplasmic reticulum (ER), trafficked to the apical plasma membrane, cleaved into the tubule and excreted in the urine, where it is the most abundant protein (Rampoldi et al., 2011). Uromodulin likely contains: four epidermal growth factor (EGF)-like domains, of which the second and third are calcium-binding (cb) EGF domains; a cysteine-rich region, which includes a domain of eight cysteines; and a zona pellucida (ZP) domain (Bokhove et al., 2016) (Fig. S1). Several functions have been postulated for uromodulin, including: regulating sodium transport in the TAL by modulating the activity of the apical transporters NKCC2 and ROMK (renal outer medullary potassium channel) (Trudu et al., 2013); inhibiting renal stone formation; preventing bacterial colonization; and modulating immune responses (Bates et al., 2004; Durisipudi et al., 2012; Mattey and Naftalin, 1992; Mo et al., 2004a,b, 2007; Raffi et al., 2009, 2005; Säämann et al., 2005a,b).

Over 90% of the >55 reported ADTKD-UMOD mutations are missense, suggesting a dominant negative disease mechanism, and >60% cause loss or gain of a cysteine residue (Moskowitz et al., 2013), implicating a role for protein misfolding in ADTKD-UMOD. Most mutations are clustered in the EGF domains and cysteine-rich region, which are likely involved in formation of most of the 24 disulfide bridges in uromodulin (Rampoldi et al., 2011), and uromodulin missense mutations cause altered intracellular trafficking, with retention of immature, core-glycosylated uromodulin in the ER, resulting in reduced or absent expression at the plasma membrane with little or no secretion (Bernascone et al., 2006; Choi et al., 2005; Dahan et al., 2003; Jennings et al., 2007; Rampoldi et al., 2003; Vylet al et al., 2006; Williams et al., 2009).
However, the role of uromodulin mutations in causing renal interstitial fibrosis, glomerulosclerosis and immune cell infiltration is not fully understood.

ER accumulation of mutant proteins can cause ER stress, which activates the unfolded protein response (UPR), which may be adaptive or apoptotic in different tissues (Tsang et al., 2010), and consists of three pathways: the activating transcription factor-6α (ATF6α), inositol-requiring enzyme-1 (IRE1) and RNA-activated protein kinase-like ER kinase (PERK) pathways. These act to upregulate chaperones and ER-associated degradation (ERAD) components, and reduce global protein translation to decrease the ER protein load. Under prolonged and unresolved ER stress, cells may also undergo apoptosis (Tsang et al., 2010). The ER stress-induced chaperone 78 kDa glucose-regulated protein [GRP78; also known as binding immunoglobulin protein (BiP)] is upregulated in renal biopsies from patients harboring mutations in UMOD (Adam et al., 2012); however, mechanisms of ER stress and the UPR pathways have not been studied further in ADTKD.

Two Umod mouse knock-out models have been developed, which have revealed roles for uromodulin in: defense against bladder and urinary tract bacterial infections (Bates et al., 2004; Mo et al., 2004b; Raffi et al., 2009, 2005); prevention of renal stone formation (Liu et al., 2010; Mo et al., 2004a, 2007); and water impermeability and solute handling in the TAL (Bachmann et al., 2005; Mutig et al., 2011; Renigunta et al., 2011; Wolf et al., 2013). However, Umod knock-out mice up to 3 years of age do not develop any of the histological features or renal failure characteristic of ADTKD (Raffi et al., 2006). A transgenic mouse model carrying a human UMOD transgene containing the ADTKD-UMOD-causing mutation Cys148Trp (C148W) in addition to the C148W mutation developed several features of ADTKD-UMOD, including mild renal failure, urinary concentrating defects, and interstitial inflammation and fibrosis, but decreased uric acid excretion was not reported (Bernascone et al., 2010) (Table S1). Thus, two similar transgenic models demonstrated two different phenotypes and, since in both models uromodulin was overexpressed compared to in non-transgenic mice, these models require cautious interpretation. An overexpressed compared to in non-transgenic mice, these models require cautious interpretation. An overexpressed compared to in non-transgenic mice, these models require cautious interpretation. An overexpressed compared to in non-transgenic mice, these models require cautious interpretation. An overexpressed compared to in non-transgenic mice, these models require cautious interpretation. An overexpressed compared to in non-transgenic mice, these models require cautious interpretation.

RESULTS

Introduction of C125R mutation into the mouse germline

A targeting vector containing 7.6 kb of the mouse Umod gene, with the C125R mutation in exon 3, a thymidine kinase (TK) cassette at the 5’ end and a neomycin resistance (NeoR) cassette in intron 2, was used to introduce the C125R mutation into the mouse genome by homologous recombination in ES cells (Fig. S1). Of 272 ES cell clones that were selection resistant, 5 clones, termed B6, C2, D11, E10 and H10, were shown to have undergone homologous recombination at the 5’ and 3’ ends of the targeting construct by PCR/restriction assay (data not shown) and Southern blot analysis (Fig. 1A). Blastocyst injection of clone C2 resulted in the birth of chimeras with a high degree of chimerism (≥90%). Male chimeras were bred with C57BL/6J females and the mutation was shown to enter the germline by genotyping of mice using PCR and BsoBI restriction digest (Fig. 1B). Germline mutant mice were bred with β-actin-Cre mice to excise the NeoR cassette. Heterozygous (Umod+/125R) and homozygous (Umod125R/125R) mutant mice were viable and fertile, with appearances and body weights (Table 1) similar to wild-type (Umod+/+) mice.

Plasma biochemistry in 8-week- and 6-month-old Umod+/+, Umod125R+/125R and Umod125R/125R mice

Male (P<0.001) and female (P<0.01) Umod125R/125R mice had plasma urea levels ~2-fold greater than those of Umod+/+ and Umod125R/125R littermates at 8 weeks of age (Table 1). By 6 months of age, plasma urea levels were increased ~1.5-fold and ~2.5-fold in Umod125R/125R (male P<0.0001; female P<0.01) and Umod125R/125R (male P<0.001; female P<0.0001) mice, respectively, compared to Umod+/+ littermates (Table 1). Plasma urea increased between 8-week- and 6-month-old Umod+/125R mice by ~1.8-fold (P<0.0001) in males and ~1.2-fold (P<0.05) in females, demonstrating progressive renal failure (Table 1). Plasma creatinine was also significantly raised, by ~1.5-fold and ~1.8-fold, in 6-month-old Umod125R/125R (P<0.05) and Umod125R/125R (P<0.001) males, respectively, and by ~1.2-fold in Umod125R/125R females (P<0.05) compared to age and sex matched Umod+/+ littermates (Table 1). Male and female 6-month-old Umod125R/125R mice also had significantly raised plasma corrected calcium concentrations (P<0.01) compared to Umod+/+ littermates, which, in conjunction with a significantly raised plasma alkaline phosphatase (ALP) activity (males P<0.001; females P<0.01), is consistent with the likely onset of tertiary hyperparathyroidism (Table 1). However, this hypercalcaemia in association with raised plasma ALP activity in the Umod125R/125R mice, which has also previously been reported in two ENU-induced mutant mouse models with A227T and C93F Umod mutations (Kemter et al., 2009, 2013) (Table S1), has not been observed to occur in ADTKD-UMOD patients who develop renal failure. A plausible explanation for this occurrence of hypercalcaemia, likely due to tertiary hyperparathyroidism, in Umod mutant mice but not ADTKD-UMOD patients is that the earlier detection of renal failure and associated secondary hyperparathyroidism, with appropriate treatment using vitamin D analogs, phosphate binders and calcimetics, will prevent the
progression to tertiary hyperparathyroidism in patients. Plasma uric acid levels were not increased in Umod+/125R or Umod125R/125R mice at 8 weeks or 6 months, likely due to expression of the hepatic enzyme uricase in mice (Wu et al., 1992). Plasma sodium and potassium concentrations were similar in mutant and wild-type mice (data not shown).

Urine biochemistry in 8-week- and 6-month-old Umod−/−, Umod+/125R and Umod125R/125R mice

Male and female Umod125R/125R mice excreted ∼2–4-fold increased volumes of urine (corrected for body weight) (P<0.0001), and this urine was more dilute with ∼2–4-fold decreased urine osmolality (8-week-old males and 6-month-old males and females P<0.0001;
Table 1. Plasma biochemistry in 8-week- and 6-month-old mice

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Male</th>
<th>Female</th>
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<tr>
<td></td>
<td>Umod&lt;sup&gt;+/+&lt;/sup&gt;</td>
<td>Umod&lt;sup&gt;+/125R&lt;/sup&gt;</td>
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<tr>
<td>8 weeks</td>
<td></td>
<td></td>
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<tr>
<td>n</td>
<td>8</td>
<td>9</td>
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<tr>
<td>Body weight (g)</td>
<td>25.8±0.3</td>
<td>26.1±0.4</td>
</tr>
<tr>
<td>Urea (mmol/l)</td>
<td>11.5±0.6</td>
<td>11.9±0.6</td>
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<tr>
<td>Creatinine (mmol/l)</td>
<td>15.5±0.9</td>
<td>16.6±0.4</td>
</tr>
<tr>
<td>Pi (mmol/l)</td>
<td>3.4±0.1</td>
<td>3.4±0.2</td>
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<tr>
<td>ALP (U/l)</td>
<td>150.9±9.5</td>
<td>143.3±5.7</td>
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<tr>
<td>Uric acid (mmol/l)</td>
<td>2.85±0.03</td>
<td>2.83±0.03</td>
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<tr>
<td>Osm (mOsm/kg)</td>
<td>345.9±3.6</td>
<td>347.4±2.6</td>
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<td>6 months</td>
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<tr>
<td>n</td>
<td>10</td>
<td>12</td>
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<tr>
<td>Body weight (g)</td>
<td>32.6±0.9</td>
<td>32.4±0.6</td>
</tr>
<tr>
<td>Urea (mmol/l)</td>
<td>13.9±0.7</td>
<td>21.5±1.4&lt;sup&gt;§&lt;/sup&gt;,&lt;sup&gt;††&lt;/sup&gt;</td>
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<tr>
<td>Creatinine (mmol/l)</td>
<td>17.6±0.9</td>
<td>25.9±4.4&lt;sup&gt;‡&lt;/sup&gt;</td>
</tr>
<tr>
<td>Pi (mmol/l)</td>
<td>2.5±0.1</td>
<td>2.8±0.1</td>
</tr>
<tr>
<td>ALP (U/l)</td>
<td>64±2.3</td>
<td>73±4.1</td>
</tr>
<tr>
<td>Uric acid (mmol/l)</td>
<td>2.6±0.02</td>
<td>2.7±0.04</td>
</tr>
<tr>
<td>Osm (mOsm/kg)</td>
<td>353±4.9</td>
<td>362±2.7</td>
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Values are mean±s.e.m.; +/+, wild types; +/125R, heterozygotes; 125R/125R, homozygotes; n denotes number of mice; ALP, alkaline phosphatase; Corr.Ca, calcium corrected for albumin; Osm, osmolality; Pi, inorganic phosphate. *P<0.05; **P<0.01; †P<0.001 compared to wild-type mice of the same age and sex. ††P<0.005; ‡‡P<0.0001 compared to 8-week-old mice of the same sex and genotype.

8-week-old females (P<0.05) compared to Umod<sup>+/+</sup> littersmates (Table 2). In addition, male Umod<sup>+/125R</sup> mice aged 8 weeks (P<0.05) and male and female Umod<sup>125R/125R</sup> mice aged 6 months (P<0.05) had reduced uric acid osmolality, and 6-month-old male Umod<sup>+/125R</sup> mice also excreted 1.5-times more urine (P<0.05) compared to Umod<sup>+/+</sup> littersmates (Table 2). In 6-month-old Umod<sup>125R/125R</sup> males and females (P<0.01), and Umod<sup>+/125R</sup> males (P<0.05), this large excretion of dilute urine caused plasma osmolality to rise and, in other mutant mice, plasma osmolalities were inappropriately normal (Table 1). Thus, Umod<sup>+/125R</sup> and Umod<sup>125R/125R</sup> mice have a urine concentrating defect, which is similar to that reported in ADTKD-UMOD patients (Rampoldi et al., 2003; Scolari et al., 2004).

Twenty-four hour uric acid excretion was decreased in 6-month-old mice by up to ∼2.4-fold in Umod<sup>+/125R</sup> mice (males P<0.05; females P<0.05), and up to ∼4.8-fold in Umod<sup>125R/125R</sup> mice (males P<0.05; females P<0.0001) compared to Umod<sup>+/+</sup> littersmates (Table 2). In 8-week-old Umod<sup>125R/125R</sup> males, 24 h uric acid excretion was also decreased by ∼1.6-fold compared to Umod<sup>+/+</sup> male littersmates (P<0.005) (Table 2). This was not due to the dilute urine produced by these mice, since the 24 h excretion takes urine volume into account. In 6-month-old mice, Umod<sup>+/125R</sup> and Umod<sup>125R/125R</sup> males and females had higher plasma inorganic phosphate and calcium corrected for albumin. In 6-month-old Umod<sup>+/125R</sup> females, plasma creatinine was decreased by ∼0.3-fold compared to Umod<sup>+/+</sup> females (P<0.005).

Table 2. Urinary parameters in 8-week- and 6-month-old mice

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Male</th>
<th>Female</th>
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<tr>
<td></td>
<td>Umod&lt;sup&gt;++&lt;/sup&gt;</td>
<td>Umod&lt;sup&gt;+/125R&lt;/sup&gt;</td>
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<tr>
<td>8 weeks</td>
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<tr>
<td>n</td>
<td>8</td>
<td>8</td>
</tr>
<tr>
<td>24 h vol/bw (µl/g)</td>
<td>99.5±5.0</td>
<td>102.3±10.7</td>
</tr>
<tr>
<td>Osm (mOsm/kg)</td>
<td>221±64</td>
<td>1960±78&lt;sup&gt;‡&lt;/sup&gt;</td>
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<tr>
<td>Cr clear (mM/min)</td>
<td>0.4±0.03</td>
<td>0.4±0.02</td>
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<tr>
<td>FEUA (%)</td>
<td>0.7±0.02</td>
<td>0.7±0.07</td>
</tr>
<tr>
<td>24 h UA (mmol)</td>
<td>115±70</td>
<td>1051±82&lt;sup&gt;‡&lt;/sup&gt;</td>
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<tr>
<td>24 h Ca (µmol)</td>
<td>3.47±0.30</td>
<td>4.12±0.47</td>
</tr>
<tr>
<td>pH</td>
<td>6.67±0.07</td>
<td>6.53±0.13</td>
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<tr>
<td>6 months</td>
<td></td>
<td></td>
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<tr>
<td>n</td>
<td>10</td>
<td>12</td>
</tr>
<tr>
<td>24 h vol/bw (µl/g)</td>
<td>76.3±7.6</td>
<td>98.6±6.7&lt;sup&gt;‡&lt;/sup&gt;</td>
</tr>
<tr>
<td>Osm (mOsm/kg)</td>
<td>1981±118</td>
<td>1198±75&lt;sup&gt;‡&lt;/sup&gt;</td>
</tr>
<tr>
<td>Cr clear (mM/min)</td>
<td>0.31±0.04</td>
<td>0.25±0.04</td>
</tr>
<tr>
<td>FEUA (%)</td>
<td>1.52±0.06</td>
<td>0.77±0.17&lt;sup&gt;‡&lt;/sup&gt;</td>
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<tr>
<td>24 h UA (mmol)</td>
<td>1024±140</td>
<td>428±47&lt;sup&gt;‡&lt;/sup&gt;</td>
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<tr>
<td>24 h Ca (µmol)</td>
<td>2.92±0.40</td>
<td>4.52±0.25&lt;sup&gt;‡&lt;/sup&gt;</td>
</tr>
<tr>
<td>pH</td>
<td>6.51±0.08</td>
<td>5.71±0.07&lt;sup&gt;‡&lt;/sup&gt;</td>
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</table>

Values are means±s.e.m.; ++, wild types; +/125R, heterozygotes; 125R/125R, homozygotes; n denotes number of mice; 24 h vol/bw, 24 h urine volume (vol) corrected for body weight (bw); Osm, osmolality; Cr clear, creatinine clearance; FEUA, fractional excretion of uric acid; 24 h UA, uric acid excretion; 24 h Ca, calcium excretion. ‡n=4 because two of the urine samples were below the lower limit of detection for uric acid concentration. ‡P<0.05; **P<0.005; ‡‡P<0.0001 compared to wild-type mice of the same age and sex.
\(Umod^{125R/125R}\) male and female mice had a significantly \((P<0.05)\) reduced FEUA, compared to \(Umod^{+/+}\) littermates (Table 2). This specific reduction in uric acid excretion in \(Umod^{+/25R}\) and \(Umod^{125R/125R}\) mice is analogous to the human ADTKD-UMOD phenotype, in which >90% of patients have a decreased FEUA (Williams et al., 2009).

Urinary calcium excretion was elevated by \(\sim 1.6\)-fold \((P<0.05)\) and \(\sim 1.8\)-fold \((P<0.005)\) in 8-week-old female \(Umod^{125R}\) and \(Umod^{125R/125R}\) mice, respectively, and elevated by \(\sim 1.5\)-fold in \(Umod^{+/125R}\) \((P<0.005)\) and \(Umod^{125R/125R}\) \((P<0.05)\) 6-month-old males, and \(\sim 1.6\)-fold \((P<0.005)\) and \(\sim 1.8\)-fold \((P<0.0001)\) in \(Umod^{125R}\) and \(Umod^{125R/125R}\) 6-month-old females, respectively, when compared to \(Umod^{+/+}\) littermates (Table 2). This hypercalciuria of \(Umod\) mutant mice, which has not been described in human ADTKD-UMOD patients, was associated with urinary acidification. Thus, urinary pH was significantly reduced in 8-week-old \(Umod^{125R/125R}\) mice, corresponding to \(\sim 1.5\)-fold \((P<0.005)\) and \(\sim 2.5\)-fold \((P<0.0001)\) increases in urinary \(H^+\) ion concentrations in males and females, respectively, compared to \(Umod^{+/+}\) littermates (Table 2). Six-month-old \(Umod^{+/125R}\) and \(Umod^{125R/125R}\) mice also had a decreased urinary \(pH\) corresponding to up to \(\sim 5.3\)-fold \((P<0.0001)\) and \(\sim 7\)-fold \((P<0.005)\) increases in urinary \(H^+\) ion concentrations in males and females, respectively, compared to \(Umod^{+/+}\) littermates (Table 2). This urinary acidification helps to prevent renal stone formation, and is a likely renal response to hypercalciuria (Renkema et al., 2009).

**Uromodulin maturation, excretion and trafficking in 8-week- and 6-month-old \(Umod^{+/125R}\) and \(Umod^{125R/125R}\) mice**

The \(C126R\) uromodulin mutation results in defective maturation, trafficking and secretion of uromodulin in vitro (Williams et al., 2009), and we therefore carried out western blot analysis of kidney lysates and urine, and immunohistochemistry, to study this in \(Umod\) mutant mice. Kidney lysates from \(Umod^{+/+}\) mice showed the presence of uromodulin at \(\sim 100\) kDa as previously described (Bates et al., 2004), corresponding to mature, fully glycosylated uromodulin (Fig. 1C). Kidney lysates from \(Umod^{+/125R}\) mice at 8 weeks and 6 months of age had mature uromodulin and a lower molecular mass form of uromodulin, likely the ER-resident precursor, which has core glycosylations only (Williams et al., 2009). Kidney lysates from \(Umod^{125R/125R}\) mice almost entirely lacked mature uromodulin, with a predominance of the precursor form of uromodulin. Higher molecular mass bands were detected in kidney lysates from \(Umod^{125R}\) and \(Umod^{125R/125R}\) mice at 8 weeks and 6 months, at approximately double the molecular mass of the precursor uromodulin; these may represent dimers of the \(Umod\) precursor, which has core glycosylations only, and 6-month-old \(Umod^{125R/125R}\) mice had a significantly decreased \(FEUA\) \((P<0.005)\) at 8 weeks of age to 15.7±2.9% at 6 months of age when compared to \(Umod^{+/+}\) littermates \((P=0.00001)\). All of the excreted uromodulin was the mature form of uromodulin at \(\sim 100\) kDa, with no detectable excretion of the precursor form of uromodulin.

Immunohistochemical analysis of uromodulin expression in 8-week- and 6-month-old \(Umod^{+/+}\), \(Umod^{125R}\) and \(Umod^{125R/125R}\) kidneys showed the expected uromodulin trafficking defect in mutant kidneys (Fig. 1E). In \(Umod^{+/+}\) kidneys, uromodulin expression had a predominantly apical pattern, with low levels of diffuse intracellular uromodulin also present as previously described (Kemter et al., 2009), and uromodulin was observed in the tubular lumen. In \(Umod^{+/125R}\) and \(Umod^{125R/125R}\) kidneys, apical expression was decreased, and dense intracellular expression was observed, such that, in \(Umod^{+/125R}\) kidneys, uromodulin was partly apical and partly intracellular, and, in \(Umod^{125R/125R}\) kidneys, uromodulin was almost exclusively located in dense intracellular deposits. Dual-labeling for uromodulin and the ER marker calnexin showed that the intracellular uromodulin in \(Umod^{+/125R}\) and \(Umod^{125R/125R}\) kidneys was colocalized with the ER (Fig. 1F), as previously demonstrated both in vitro (Williams et al., 2009) and in ADTKD-UMOD patients (Bernacone et al., 2010), and consistent with the presence of the lower molecular mass precursor detected by western blotting (Fig. 1C). In \(Umod^{125R}\) mouse kidneys, uromodulin colocalized with calnexin and was present at the apical membrane.

**Renal fibrosis and immune cell infiltration in 8-week- and 6-month-old \(Umod^{+/125R}\) and \(Umod^{125R/125R}\) mice**

Masson’s trichrome staining, to study renal damage and fibrosis, of kidney sections from 8-week- and 6-month-old mice revealed that at 8 weeks, kidneys of \(Umod^{+/125R}\) mice did not have any abnormalities, but that kidneys of \(Umod^{125R/125R}\) mice had thickened basement membranes and some intratubular fibrous deposits, similar to those reported in a renal biopsy from a patient with a lower \(UMOD\) mutation (Nasar et al., 2008). Glomeruli appeared normal in both \(Umod^{+/125R}\) and \(Umod^{125R/125R}\) mouse kidneys at 8 weeks of age (Fig. 2A). At 6 months of age, \(Umod^{+/125R}\) mouse kidneys had mild interstitial fibrosis, and \(Umod^{125R/125R}\) mouse kidneys had large areas of fibrosis, with some renal tubules appearing to become detached from the basement membrane. Furthermore, some glomeruli in \(Umod^{125R/125R}\) mouse kidneys also had increased fibrotic material, consistent with glomerulosclerosis (Fig. 2A). No cysts were observed in \(Umod^{125R}\) or \(Umod^{125R/125R}\) mouse kidneys at 8 weeks or 6 months of age.

During analysis of uromodulin-labeled and trichrome-stained kidney sections, interstitial cell infiltrates were noted. To determine the nature of these infiltrates, kidney sections were labeled for T-cell (CD3) and macrophage (F4/80) markers. \(Umod^{125R}\) mouse kidneys at 8 weeks of age had no detectable immune cell infiltration; however, \(Umod^{125R/125R}\) mouse kidneys showed small areas of interstitial infiltrates, consisting of T-cells and macrophages (Fig. 2B,C). At 6 months of age, \(Umod^{125R}\) mouse kidneys showed small areas of interstitial T-cells and macrophages, but \(Umod^{125R/125R}\) mouse kidneys had multiple, large areas of interstitial infiltrates, consisting of both T-cells and macrophages (Fig. 2B,C). These infiltrates occurred mainly in the medullary region, and often in close proximity to TAL cells expressing mutant uromodulin.
Upregulation of ER stress and UPR pathways in mutant TALs

Accumulation of mutant proteins in the ER may cause ER stress, triggering an adaptive pathway called the unfolded protein response (UPR). Members of the chaperone family of GRPs, and in particular GRP78, are upregulated by the UPR in response to ER stress. We therefore used immunohistochemistry of serial kidney sections to investigate GRP78 expression in TAL cells expressing mutant uromodulin. In Umod<sup>+/+</sup> kidneys, GRP78 was expressed at a very low level in non-TAL tubular epithelial cells, and at a slightly elevated level in TAL tubular cells (Fig. 3A,B), a finding that was confirmed in kidneys from an unrelated mouse line (data not shown). However, in kidneys from both 8-week- and 6-month-old Umod<sup>+/125R</sup> and Umod<sup>125R/125R</sup> mice, GRP78 expression was upregulated specifically in TAL cells expressing mutant uromodulin, and not in other tubular segments (Fig. 3A,B).

We sought to determine which of the three UPR pathways may be active in TAL cells expressing mutant uromodulin, by immunohistochemistry to detect expression of a member of each of the three pathway, which are: ATF6α (ATF6α pathway), spliced (active) XBP1 (XBP1<sup>S</sup>; IRE1 pathway) or ATF4 (PERK pathway). None of the three pathways were upregulated in TAL cells in Umod<sup>+/+</sup> mice when compared to non-TAL cells (Fig. 4A,B). No activation of the ATF6α pathway was detected in Umod<sup>+/125R</sup> or Umod<sup>125R/125R</sup> mice at either 8 weeks or 6 months of age, as there was no increase in nuclear ATF6α expression in TAL cells (Fig. 4A,B). By contrast, both XBP1<sup>S</sup> and ATF4 were upregulated in TAL cells of Umod<sup>+/125R</sup> and Umod<sup>125R/125R</sup> mice compared to Umod<sup>+/+</sup> littermates at both 8 weeks and 6 months of age (Fig. 4A,B), demonstrating upregulation of the IRE1 and PERK pathways, respectively, in response to retention of mutant uromodulin in the ER.

In order to quantify the upregulation of these ER stress and UPR pathways, we microdissected TALs from Umod<sup>+/+</sup> and Umod<sup>125R/125R</sup> mice. Quantitative real-time PCR (qRT-PCR) analysis for nephron segment-specific genes was undertaken to demonstrate the purity of the obtained TALs (data not shown), followed by qRT-PCR and western blot analyses for UPR components. qRT-PCR
analysis showed that Xbp1S and Atf4 were significantly upregulated by ∼1.9-fold (P<0.01) and ∼1.6-fold (P<0.001), respectively, in Umod125R/125R TALs compared to Umod+/+ TALs, but that Grp78 was significantly downregulated by ∼4.2-fold (P<0.001) in Umod125R/125R TALs compared to Umod+/+ TALs, consistent with regulation of GRP78 expression at the translational level (Gulow et al., 2002) (Fig. 5A). Interestingly, ATF6α was significantly downregulated by ∼1.8-fold (P<0.05) in Umod125R/125R TALs compared to Umod+/+ TALs (Fig. 5B,C). XBP1S and ATF4 were significantly upregulated by ∼13.6-fold (P<0.001) and ∼4.5-fold (P<0.01), respectively, in Umod125R/125R TALs (Fig. 5B,C), confirming the results obtained by immunohistochemical analysis.

Upregulation of the PERK pathway via ATF4 is associated with suppression of general protein translation but induction of expression of specific UPR target genes, such as CCAAT/enhancer-binding protein homologous protein (CHOP) (Hetz, 2012). qRT-PCR analysis of isolated TALs showed that Umod mRNA was downregulated by ∼6.6-fold (P<0.001) in Umod125R/125R TALs compared to Umod+/+ TALs (Fig. 5A). Chop mRNA was upregulated by ∼2.2-fold (P<0.001) in Umod125R/125R TALs compared to Umod+/+ TALs (Fig. 5A), and immunohistochemical analysis of serial kidney sections confirmed that CHOP protein was upregulated in TAL cells in Umod+/+ and Umod125R/125R mice compared to Umod+/+ littermates at both 8 weeks and 6 months of age, as shown by accumulation of CHOP in the nuclei of TAL cells (Fig. 6A,B). Increased CHOP expression may lead to apoptosis (Hetz, 2012); however, analysis of kidney sections using terminal deoxynucleotidyl transferase dUTP nick end labeling (TUNEL) assay and co-staining for uromodulin showed that, as in Umod+/+ littermates, there were no TUNEL-positive TAL cells in Umod+/+ and Umod125R/125R mouse kidneys at either 8 weeks or 6 months of age (Fig. 6A,B). However, occasional TUNEL-positive cells were detected in other segments, demonstrating that the assay was functional. Thus, mechanisms other than apoptosis are likely responsible for tubular damage in Umod+/+ and Umod125R/125R mice.

**DISCUSSION**

We have generated a mouse model with an ADTKD disease-causing UMOD mutation, that displays: defective uric acid excretion; urinary concentrating defects; renal failure; defective uromodulin trafficking, maturation and secretion; renal fibrosis; interstitial immune cell infiltration; and ER stress with upregulation of the UPR in the TAL. This model is representative of ADTKD-UMOD patients, who develop hyperuricemia, low FEUA and gout, chronic renal failure, tubulointerstitial nephropathy, glomerulosclerosis and lymphocytic infiltration (Lhotta et al., 1998; Turner et al., 2003). Thus, this mouse provides an in vivo model to study mechanisms of renal fibrosis and ER stress.

A major feature of ADTKD-UMOD in humans is hyperuricemia, which was not present in Umod mutant mice. This is not surprising, as mice express the hepatic enzyme uricase (urate oxidase), which catalyses low solubility uric acid to the high solubility allantoin; uricase has been evolutionarily silenced in humans and other primates (Wu et al., 1992). However, Umod+/+ and Umod125R/125R mice, similar to ADTKD-UMOD patients, still demonstrated significant decreases in both FEUA and 24 hour uric acid excretion. Interestingly, in female Umod125R/125R mice, this decreased uric acid excretion was detected after the onset of renal failure, as measured by a significantly increased plasma urea, since 8-week-old Umod125R/125R females had an elevated plasma urea, but a similar uric acid excretion compared to Umod+/+ females. The role of plasma uric acid levels in the pathogenesis of ADTKD-UMOD has been the subject of much debate, particularly whether drugs that lower plasma uric acid can slow the progression of ADTKD-UMOD in patients (Bleyer and Hart, 2003; Fairbanks et al., 2002, 2004; Puig et al., 2006; Puig and Torres, 2003, 2004).
Our model demonstrates that hyperuricemia is a secondary consequence of the other phenotypes of ADTKD-UMOD (e.g. decreased uric acid excretion, renal failure and kidney histopathology), and is not required to cause the renal pathophysiology, since our model develops these other phenotypes without hyperuricemia. However, hyperuricemia may help to propagate or enhance the disease progression in humans, and this hypothesis could be tested in our model by using uricase inhibitors.

The mechanism of decreased uric acid excretion is not fully understood, but may be a consequence of the urine concentrating defect. In the normal TAL, the water impermeability is likely due to the presence of uromodulin in a gel-like structure at the apical plasma membrane, which is permeable to ions but not to water (Mattey and Naftalin, 1992). In Umod^{+/125R} and Umod^{125R/125R} mice, retention of the mutant uromodulin protein in the ER of the TAL cells would decrease the amount of uromodulin at the apical plasma membrane, and may compromise this water impermeability. This may lead to a reduction in the normally high osmolality of the medullary interstitium, thus inhibiting water reabsorption in the collecting duct (CD), and resulting in a more dilute urine (Devuyst et al., 2005). The previously reported Umod^{−/−} knock-out mice also have a urine concentrating defect (Bachmann et al., 2005). This defect in urine concentration due to alterations of TAL characteristics may be analogous to patients treated with loop diuretics, such as those with hypertension, who also develop secondary hyperuricemia (Ramsay et al., 1994). Loop diuretics (e.g. furosemide) specifically inhibit the TAL Na+:K+:2Cl− (NKCC2) transporter, decreasing water reabsorption with a consequent increase in urine volume (Gamba and Friedman, 2009). During loop diuretic treatment, the resulting decrease in extracellular fluid (ECF) volume causes increased sodium reabsorption by the proximal convoluted tubule (PCT), which likely drives increased uric acid reabsorption, hence causing the hyperuricemia which is often associated with these drugs (Weinman et al., 1975). A possible compensatory mechanism for excess water loss has been described in Umod^{−/−} knock-out mice by upregulation of several critical TAL ion transporters, including the target of loop diuretics, NKCC2, which is usually activated by uromodulin (Bachmann et al., 2005). Thus, similar mechanisms may
be responsible for hyperuricemia in loop diuretic-treated hypertensive patients and ADTKD-UMOD patients; this requires further investigation in Umod^{+/125R} and Umod^{125R/125R} mice.

The urine concentrating defect described in Umod^{-/-} knock-out mice (Bachmann et al., 2005) and Umod^{+/125R} and Umod^{125R/125R} mice in this study is likely due to the absence of uromodulin at the apical plasma membrane of the TAL. However, Umod^{-/-} knock-out mice do not develop any histological characteristics of ADTKD-UMOD, or renal failure (Raffi et al., 2006). Thus, mutant uromodulin protein must be present to cause the renal pathology associated with ADTKD-UMOD, and we have shown that histopathological changes characteristic of human ADTKD-UMOD patients, such as interstitial fibrosis and immune cell infiltration, are present in our Umod^{-/-} and Umod^{125R/125R} knock-in mice. Uromodulin is the most abundant protein in the urine, and is expressed only by the TAL (Scolari et al., 2015), indicating a high rate of uromodulin expression in TAL cells. Thus, even though cells possess mechanisms to degrade misfolded proteins, such as ERAD, the high expression rate of mutant uromodulin over time may overwhelm such mechanisms. Uromodulin can also stimulate the innate and adaptive immune systems (Säemann et al., 2005b; Su et al., 1997). Thus, it has been postulated that kidney damage may compromise its integrity and allow uromodulin to be exposed to the immune system, which may cause antibody production (Säemann et al., 2005b), as well as inflammation and/or immune cell infiltration, which can result in fibrosis of the kidney (Eddy, 2014).

Umod^{+/125R} and Umod^{125R/125R} mice have several phenotypic characteristics that are similar to those reported for the C147W uromodulin transgenic (Tg^{UmodC147W}) mouse model, including renal failure, urinary concentrating defects, fibrosis, immune cell infiltration and intracellular uromodulin retention. However, Umod^{+/125R} and Umod^{125R/125R} mice had a selective defect in renal uric acid handling that was not studied in Tg^{UmodC147W} transgenic mice (Bernascone et al., 2010), whereas Tg^{UmodC147W} mice had renal cysts, which were not detected in Umod^{+/125R} or Umod^{125R/125R} mice (Table S1). The reason for these differences between Umod^{125R} mice and Tg^{UmodC147W} mice remains to be elucidated, but there are several possible explanations. First, these differences may involve strain-specific factors, as the Umod^{125R} mice were on a C57BL/6 background, whilst the Umod^{125R/125R} mice were on a FVB background, whilst Umod^{C147W} mice were generated on an FVB background, whilst the Umod^{125R/125R} mice had a selective defect in renal uric acid handling that was not studied in Tg^{UmodC147W} transgenic mice (Bernascone et al., 2010), whereas Tg^{UmodC147W} mice had renal cysts, which were not detected in Umod^{+/125R} or Umod^{125R/125R} mice (Table S1). The reason for these differences between Umod^{125R} mice and Tg^{UmodC147W} mice remains to be elucidated, but there are several possible explanations. First, these differences may involve strain-specific factors, as the Umod^{125R} mice were on a C57BL/6 background, whilst the Umod^{125R/125R} mice were on a FVB background, whilst the Umod^{C147W} mice were generated on an FVB background, whilst the Umod^{125R/125R} mice had a selective defect in renal uric acid handling that was not studied in Tg^{UmodC147W} transgenic mice (Bernascone et al., 2010), whereas Tg^{UmodC147W} mice had renal cysts, which were not detected in Umod^{+/125R} or Umod^{125R/125R} mice (Table S1). The reason for these differences between Umod^{125R} mice and Tg^{UmodC147W} mice remains to be elucidated, but there are several possible explanations. First, these differences may involve strain-specific factors, as the Umod^{125R} mice were on a C57BL/6 background, whilst the Umod^{125R/125R} mice were on a FVB background, whilst the Umod^{C147W} mice were generated on an FVB background, whilst the Umod^{125R/125R} mice had a selective defect in renal uric acid handling that was not studied in Tg^{UmodC147W} transgenic mice (Bernascone et al., 2010), whereas Tg^{UmodC147W} mice had renal cysts, which were not detected in Umod^{+/125R} or Umod^{125R/125R} mice (Table S1). The reason for these differences between Umod^{125R} mice and Tg^{UmodC147W} mice remains to be elucidated, but there are several possible explanations. First, these differences may involve strain-specific factors, as the Umod^{125R} mice were on a C57BL/6 background, whilst the Umod^{125R/125R} mice were on a FVB background, whilst the Umod^{C147W} mice were generated on an FVB background, whilst
families with either the C126R or C148W mutations (Lhotta et al., 1998; Rampoldi et al., 2003), and the basis for this variation in cyst formation between different families, family members and mouse models requires further investigation. Two ENU-mutagenized mouse models, carrying A227T or C93F uromodulin mutations on the same backgrounds, have recently been described to have differing rates of disease onset and progression, with C93F mutant mice having the more severe phenotype (Kemter et al., 2013). The ENU mutant mice both displayed decreased uric acid excretion that was present in \textit{Umod}^{125R} mice but absent in \textit{Tg}^{UmodC147W} mice, tubulocystic changes that were present in \textit{Tg}^{UmodC147W} mice but absent in \textit{Umod}^{125R} mice, and glomerulocystic changes that were absent in both \textit{Tg}^{UmodC147W} and \textit{Umod}^{125R} mice (Bernascone et al., 2010; Kemter et al., 2013) (Table S1). These differences between the germline \textit{Umod}^{125R}, transgenic \textit{Tg}^{UmodC147W} and ENU A227T and C93F mice with different mutations highlight the advantages of having multiple mouse models that provide the opportunity of elucidating further the relationships between protein, uromodulin mutation, genetic modifiers, phenotype and mechanism in ADTKD-\textit{UMOD}.

Our results provide mechanistic insights by showing that ER stress is present in TAL cells expressing mutant uromodulin, and that two of the three UPR pathways (PERK and IRE1) are active, leading to upregulation of CHOP. However, this did not lead to apoptosis of TAL cells, demonstrating that other mechanisms may be responsible for TAL damage, and consequent fibrosis and immune cell infiltration, in this model. CHOP is a transcription factor typically thought to upregulate pro-apoptotic pathways; however, it may also be protective under certain conditions in different cell types. For example, mice with autosomal dominant retinitis pigmentosa due to ER retention of mutant rhodopsin have retinal damage that is associated with UPR and CHOP upregulation; however, deletion of CHOP led to increased retinal damage in these mice, suggesting a protective role for CHOP (Nashine et al., 2013).

Several studies have implicated ER stress, leading to CHOP expression and apoptosis, in models of renal disease. However, these studies have primarily investigated the renal response to toxins (e.g. high glucose in diabetic nephropathy or high albumin levels to mimic proteinuria), drugs (e.g. acetaminophen) or acute renal injury (e.g. using unilateral ureteric obstruction), or have focused on glomerular injury (Dickhout and Krepinsky, 2009). Relatively little work has been carried out to understand the role of chronic ER stress in renal tubule cells endogenously expressing an ER-retained mutant protein, and our mouse model for ADTKD-\textit{UMOD} will be an important resource for further studies in this area. Active XBP1 protein was also upregulated in TAL cells in \textit{Umod}^{125R} mice,
demonstrating upregulation of the IRE1 pathway, and this may also be an adaptive or apoptotic response to ER stress (Ghosh et al., 2014). During prolonged ER stress, PERK signaling is maintained whilst IRE1 signaling is attenuated, which may lead to a switch from adaptive to apoptotic signaling (Lin et al., 2007). Consistent with this hypothesis, prolonged IRE1 signaling enhanced cell proliferation and prevented apoptosis when human embryonic kidney (HEK293) cells were exposed to the UPR inducers tunicamycin and thapsigargin (Lin et al., 2007, 2009). In *Umod*<sup>125R</sup> mice, both the PERK and IRE1 pathways appeared to be active even at 6 months of age, and the presence of both of these pathways may explain the absence of the apoptotic response in TAL cells in these mice. Studies of the transgenic Tg<sup>UmodCRE125R</sup> mice also showed an absence of apoptosis by a lack of active caspase-3 staining; thus, apoptosis does not appear to play a significant role in tubule damage in ADTKD-*UMOD* (Bernascone et al., 2010).

In conclusion, we have generated a mouse model for a monogenic cause of renal fibrosis carrying an ADTKD-*UMOD* disease-causing mutation, C125R, in the endogenous mouse *Umod* gene. *Umod<sup>125R</sup>* and *Umod<sup>125R</sup>+125R* mice demonstrate several of the phenotypes of human ADTKD-*UMOD* patients, including progressive renal failure, urine concentrating defects, decreased uric acid and uromodulin excretion, retention of uromodulin in the ER, and pathological changes in the kidney such as fibrosis, immune cell infiltration and ER stress. The PERK and IRE1 UPR pathways were found to be specifically upregulated in TAL cells of *Umod<sup>125R</sup>* and *Umod<sup>125R</sup>+125R* mice, which was associated with upregulation of CHOP; however, this did not lead to apoptosis. This novel knock-in mouse model will be a valuable resource to study the mechanisms of pathogenesis and progression of renal fibrosis, ESRF and ADTKD-*UMOD*, and to test novel therapeutic agents for these diseases.

**MATERIALS AND METHODS**

**Generation of knock-in targeting vector and mutant mice**

Exon 3 of mouse *Umod* was amplified by PCR and used to screen a mouse PAC library (MRC Geneservice, Cambridge, UK). A PAC containing mouse *Umod* was then used as a template in a long-range PCR reaction to amplify an ∼7.6 kb product between the 5′ upstream region and intron 5 of *Umod*. exon and intron-exon boundary sequences were verified by direct DNA sequencing as described (Nesbit et al., 2004). After insertion into pGEM-T (Promega, Southampton, UK), site-directed mutagenesis (SDM) was performed to introduce the C125R mutation as described (Nesbit et al., 2004); during this procedure, a novel BsoBI restriction site was also generated. SDM was also used to introduce a HindIII site into intron 2. Following each round of SDM, the sequences of exons and intron-exon boundaries were verified by direct DNA sequencing. A neomycin resistance (*neo*) cassette flanked by two *loxP* sites was excised from plasmid pmmneofox8 and cloned into intron 2 of *Umod* using XbaI and HindIII. Plasmid pBSTK, containing a thymidine kinase (TK) cassette, was mutagenized by SDM to introduce a *SfiI* restriction site, and the TK cassette was inserted into the targeting vector at the 5′ end of the *Umod* gene using *SfiI* and *SacI* (Fig. S1). The vector was linearized using *NotI*, electroporated into the R1 embryonic stem (ES) cell line, and cells selected using 300 µg/ml G418 (Geneticin, Gibco, Paisley, UK) and 0.2 µM gancyclovir (Calbiochem, Manchester, UK). Cells in which homologous recombination had occurred at both the 5′ and 3′ arms were identified by PCR/restriction digest and Southern blot analysis (Harding et al., 2009) as follows: 5′ PCR: forward primer in the 5′ region outside the vector and reverse primer within the Neo<sup>R</sup> cassette generated a 2.8 kb product in recombined clones only; 3′ PCR: forward primer within exon 3 and reverse primer within intron 5 (outside the vector) generated a 5.3 kb fragment in all clones, which, when digested with BsoBI, generated an extra fragment in recombined clones; 5′ Southern blot: digestion with *PstI* and a probe within the 5′ upstream region detected an ∼11.6 kb *Umod* wild-type fragment and an ∼7.4 kb *Umod* mutant fragment; 3′ Southern blot: digestion with SpeI and a probe corresponding to exon 6 of *Umod* detected an ∼8.3 kb *Umod* wild-type fragment and an ∼9.6 kb *Umod* mutant fragment (Fig. 1A). ES cells heterozygous for the mutation were injected into C57BL/6 blastocysts at embryonic day 2.5, which were transferred into the uterus of pseudopregnant C57BL/6 females. Male chimeras were mated with C57BL/6 females and the offspring tested for germline transmission by extraction of DNA from ear biopsy material in DNA extraction buffer (50 mM Tris pH 8.5, 1 mM EDTA, 0.5% Tween-20, with 50 µg Proteinase K), followed by PCR and BsoBI digestion. The Neo<sup>R</sup> cassette was excised between *loxP* sites by mating with β-actin-Cre mice, and the removal of the Neo<sup>R</sup> cassette was verified by PCR assay. Mice were backcrossed to the C57BL/6J background strain and mice from backcross (BC3)-BC5 were used for studies in 6-month-old mice, and from BC4-BC7 for studies in 8-week-old mice. Animals were maintained according to local welfare guidelines, and all procedures were carried out under UK Home Office personal and project license restrictions.

**Metabolic cage analysis**

Eight-week- and 6-month-old *Umod<sup>+/+</sup>, *Umod<sup>+125R/R</sup>* and *Umod<sup>125R/125R</sup>* littermates were investigated in individual metabolic cages (Techniplast) for 24 h with free access to food and water. Body weight, water intake and urine output were measured and urine was collected under mineral oil. Mice were euthanized by terminal anesthesia, blood collected via the jugular vein, and plasma obtained by centrifugation at 800 g for 10 min. Plasma and urine biochemical analysis was performed on a Beckman Coulter AU680 semi-automated clinical biochemistry analyzer as described (Piret et al., 2012), and plasma calcium was corrected for albumin (Corr.Ca) using the formula: Corr.Ca=Ca/(1−0.017) (Piret et al., 2012). Osmolalities were measured using a Model 3320 Osmometer (Advanced Instruments Inc., Norwood, MA, USA) and urinary pH was measured using a Microprocessor pH meter (Hanna Instruments, Leighton Buzzard, UK).

**Western blot analysis of whole kidney lysates and urine**

Uromodulin maturation and excretion in *Umod<sup>+/+</sup>, *Umod<sup>+125R/R</sup>* and *Umod<sup>125R/125R</sup>* mutant mice were studied by western blot analysis of kidney lysates and urine samples. Kidney samples were homogenized in β-octylglucoside lysis buffer [20 mM Tris-HCl pH 8; 150 mM NaCl; 60 mM β-octyl-glucoside (Bernascone et al., 2010)] with a protease inhibitor cocktail (Roche, Burgess Hill, UK). Insoluble material was removed by centrifugation, total protein concentration of the soluble fraction determined using the micro BCA Protein Assay (Pierce, UK) (Bernascone et al., 2010; Williams et al., 2009), and lysates loaded at 20-50 µg total protein per lane. Urine samples were normalized to urinary creatinine such that, for *Umod<sup>+/+</sup>* and *Umod<sup>+125R/R</sup>* samples, a urine volume corresponding to 0.5 mmol creatinine was loaded and, for *Umod<sup>125R/125R</sup>* samples, a urine volume corresponding to 10 mmol was loaded. Western blotting and densitometric analysis were performed as previously described, using sheep anti-uromodulin antibody (Biodiesation International, Waco, ME, USA), or rabbit anti-tubulin antibody (Abcam ab15246, 1:500), followed by horseradish peroxidase (HRP)-conjugated mouse anti-sheep (Sigma, Poole, UK) or HRP-conjugated goat anti-rabbit (Bio-Rad, Watford, UK) antibodies, respectively (Williams et al., 2009). To control for variances between blots of urinary uromodulin, at least two *Umod<sup>+/-</sup>* samples were loaded on each gel, a standard ‘remove background’ tool was used prior to quantification by densitometry, and mean densities of *Umod<sup>-/-</sup>* bands were set to 100% for comparison with other bands on the same blot.

**Histological and immunohistochemical analysis**

Kidneys were dissected and fixed in 10% neutral buffered formalin for 24 h, then dehydrated, paraffin embedded and sectioned at 5 µm thickness. Trichrome staining was carried out according to standard protocols. For immunohistochemical labeling, sections were rehydrated, then subjected to antigen unmasking either by incubation in 20 µg/ml proteinase K for 5-15 min, or by heat-mediated antigen retrieval in either pH 6, pH 9 or pH −5.15 min, or by heat-mediated antigen retrieval in either pH 6, pH 9 or pH 9.5. After antigen unmasking, sections were rehydrated, stained with a standard antibody cocktail (Roche) and then subjected to immunohistochemical labeling. Sections were rehydrated, subjected to antigen unmasking either by incubation in 20 µg/ml proteinase K for 5-15 min, or by heat-mediated antigen retrieval in either pH 6, pH 9 or pH 9.5. After antigen unmasking, sections were rehydrated, stained with a standard antibody cocktail (Roche) and then subjected to immunohistochemical labeling.
10 buffers. Sections were blocked in pre-immune serum, and probed with the following primary antibodies: sheep anti-uromucoid (Biodesign International, 1:500); rabbit anti-calcxin (Abcam ab22595, 1:50); rabbit anti-CD3 (Abcam ab16669, 1:50); rat anti-F4/80 (Serotec clone A3-1, 1:50); rabbit anti-GRP78 (Abcam ab21685, 1:1000); rabbit anti-ATF4 (Abcam ab105383, 1:500); rabbit anti-ATF6α [Santa Cruz Biotechnology (Santa Cruz, CA, USA) sc-22799, 1:50]; goat anti-XBP1 (spliced) (Abcam ab85546, 1:100); or rabbit anti-CHOP (Santa Cruz sc-575, 1:50). Secondary detection was achieved by: HRP-conjugated donkey anti-sheep antibody (Jackson ImmunoResearch, 1:500) visualized using the 3,3′-Diaminobenzidine (DAB) reaction kit (Vector Labs, Peterborough, UK) for uromodulin; the DAKO Envision+ System DAB kit for rabbit primary antibodies; the Santa Cruz ABC staining system for goat primary antibodies; or fluorescently-conjugated donkey secondary antibodies (Jackson ImmunoResearch or Invitrogen) for dual-labeling. TUNEL assay was performed using the ApopTag Fluorescein kit (Millipore, Hertfordshire, UK) according to the manufacturer’s instructions. DAB-stained slides were counterstained with hematoxylin, and fluorescently labeled slides were counterstained with 4′,6-diamidino-2-phenylindole (DAPI). Slides were examined by light or UV microscopy using an Eclipse E400 microscope (Nikon, Japan), and images were captured using a DXM1200C digital camera, with an original magnification of 20× or 40×, and NIS-Elements BR2.30 software (both Nikon). Immunohistochemistry was performed on 2 kidney sections from each of 4 mice per group.

**Isolation of mouse TALs, and qRT-PCR and western blot analyses**

Mouse TALs were isolated following a previously described protocol (Glaudemans et al., 2014) with minor modifications. Briefly, 2- to 3-month-old male Umod+/+ and Umod125R/125R mice were sacrificed by cervical dislocation. Kidneys were removed, decapsulated and placed in ice-cold dissection solution (Hanks balanced salt solution with 15 mM HEPES, 10 mM D-glucose, 5 mM glycine, 1 mM L-alanine, adjusted to 325 mOsm/kgH2O with mannitol and pH 7.4 with NaOH). Each kidney was cut into 2 equal parts by a midsagittal section and then into thin transverse slices that were further cut into radial corticomedullary stripes. Kidney pieces were digested for 30 min in dissection solution supplemented with 245 U/ml type 2 collagenase (Worthington Biochemical Corporation, Lakewood, NJ, USA) and 96 µg/ml soybean trypsin inhibitor (Sigma-Aldrich) at 37°C on a shaker. The supernatant containing small kidney fragments and isolated tubules was sieved through 250 µm and 50 µm filters (Sefar AG, Heiden, Switzerland) and tubules retained on the 50 µm filter were collected in 37°C dissection solution supplemented with 1% BSA (Sigma-Aldrich) and 96 µg/ml soybean trypsin inhibitor (Sigma-Aldrich). Remaining undigested kidney fragments were digested for 10 min in fresh collagenase solution. TALs were manually selected under a light microscope using a glass pipet connected to a micromanipulator (Narishige International, London, UK). Total RNA was extracted from microdissected TALs using RNaseq® Total RNA Isolation Kit (Invitrogen, MA, USA), following the manufacturer’s protocol, and DNase1 treatment performed. The reverse transcriptase (RT) reaction was performed using 1 µg of RNA and the iScript™ cDNA Synthesis Kit (Bio-Rad). Changes in mRNA levels of target genes were determined by relative qRT-PCR with a CFX96™ Real-Time PCR Detection System (Bio-Rad) using iQ™ SYBR Green Supermix (Bio-Rad) detection of single PCR product accumulation in duplicate. Specific primers were designed using Primer3. PCR conditions were 95°C for 3 min followed by 40 cycles of 15 s at 95°C, 30 s at 60°C. PCR products were sequenced using the BigDye Terminator Kit (Perkin Elmer Applied Biosystems, MA, USA), purified using the multiScreen SEQ384 Filter Plate (Millipore) and Sephadex G-50 DNA Grade Fine (Amer sham Biosciences, NJ, USA), and analyzed on an ABI1300 capillary sequencer (Perkin Elmer Applied Biosystems). The efficiency of each set of primers was determined by dilution curves. Glyceraldehyde 3-phosphate dehydrogenase (Gapdh) was used routinely as a reference gene. Changes in relative target gene/Gapdh mRNA ratio were determined by the formula: 2ΔΔCt. For western blot analysis, 50-70 TALs were pooled and directly transferred in 4× Laemmli sample buffer (Bio-Rad supplemented with protease inhibitors (Roche, Basel, Switzerland) and 60 µg/ml diithiothreitol (AppliChem), and boiled at 95°C for 5 min. Western blotting and densitometric analysis were performed using antibodies against ATF4, ATF6α, GRP78 and spliced XBP1, as above.

**Statistical analysis**

Statistical analyses were performed using Student’s unpaired t-test or the Mann–Whitney U-test, where appropriate.

**Study approval**

All animal studies were approved by the University of Oxford Ethical Review Committee and were licensed under the Animals (Scientific Procedures) Act 1986, issued by the United Kingdom Government Home Office Department.

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**Competing interests**

The authors declare no competing or financial interests.

**Author contributions**


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**Supplementary information**

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