A novel mouse model of anterior segment dysgenesis (ASD): conditional deletion of Tsc1 disrupts ciliary body and iris development

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ABSTRACT
Development of the cornea, lens, ciliary body and iris within the anterior segment of the eye involves coordinated interaction between cells originating from the ciliary margin of the optic cup, the overlying periccular mesenchyme and the lens epithelium. Anterior segment dysgenesis (ASD) encompasses a spectrum of developmental syndromes that affect these anterior segment tissues. ASD conditions arise as a result of dominantly inherited genetic mutations and result in both ocular-specific and systemic forms of dysgenesis that are best exemplified by aniridia and Axenfeld–Rieger syndrome, respectively. Extensive clinical overlap in disease presentation amongst ASD syndromes creates challenges for correct diagnosis and classification. The use of animal models has therefore proved to be a robust approach for unravelling this complex genotypic and phenotypic heterogeneity. However, despite these successes, it is clear that additional genes that underlie several ASD syndromes remain unidentified. Here, we report the characterisation of a novel mouse model of ASD. Conditional deletion of Tsc1 during eye development leads to a premature upregulation of mTORC1 activity within the ciliary margin, pericellular mesenchyme and lens epithelium. This aberrant mTORC1 signalling within the ciliary margin in particular leads to a reduction in the number of cells that express Pax6, Bmp4 and Msx1. Sustained mTORC1 signalling also induces a decrease in ciliary margin progenitor cell proliferation and a consequent failure of ciliary body and iris development in postnatal animals. Our study therefore identifies Tsc1 as a novel candidate ASD gene. Furthermore, the Tsc1-ablated mouse model also provides a valuable resource for future studies concerning the molecular mechanisms underlying ASD and acts as a platform for evaluating therapeutic approaches for the treatment of visual disorders.

KEY WORDS: Tsc1, mTORC1, Pax6, Ciliary body, Iris, Anterior segment dysgenesis

INTRODUCTION
Anterior segment dysgenesis (ASD) encompasses a spectrum of autosomal dominant developmental syndromes that affect tissues within the anterior segment of the eye including the cornea, lens, iris, ciliary body (CB) and associated drainage structures involving the trabecular meshwork (TM) and Schlemm’s canal. Development of these tissues involves a complex interplay between neuroectodermal cells originating from the ciliary margin (CM) of the optic cup, neural crest cells residing within the overlying pericellular mesenchyme (POM) and ectodermal lens epithelial (LE) cells. ASD is caused by genetic mutations that affect these cellular domains and presents as either autonomous ocular anomalies or ocular anomalies accompanied by systemic deficits. Ocular-specific ASD is a group of syndromes that solely affect the eye and its associated structures. This group is best exemplified by aniridia which is a neuroectoderm-derived panocular disorder that is associated predominantly with mutations in paired box gene 6 (PAX6). Individuals affected by aniridia present with distinct anterior segment deficits such as iris hypoplasia and cataracts. By contrast, the systemic forms of ASD are a group of closely related diseases with Axenfeld–Rieger syndrome (ARS) and Peter’s anomaly (PA) being amongst the best characterised. Both are neural crest-derived disorders and result predominantly from mutations in either paired-like homeodomain 2 (PITX2) or forkhead box C1 (FOXC1). ARS and PA share common and highly penetrant anterior segment deficits that primarily affect the pupil and drainage structures (Cvekl and Tamm, 2004; Gould and John, 2002; Graw, 2010; Idrees et al., 2006; Ito and Walter, 2014; Lee et al., 2008; Reis and Semina, 2011; Sowden, 2007).

Diagnosis of the precise ASD syndrome is challenging because of the extensive clinical overlap observed amongst related diseases. The generation of animal models has therefore proved to be an effective approach for deciphering the molecular aetiology underlying this heterogeneity (Gould and John, 2002). Haploinsufficiency in the mouse Pax6 gene results in a clinical model of aniridia with iris hypoplasia being prevalent; while modulation of Pitx2 and Foxc1 gene dose results in pupil and drainage structure abnormalities and thus represent experimental models of ARS (Baulmann et al., 2002; Gage et al., 1999; Hill et al., 1991; Hogan et al., 1988; Holmberg et al., 2004; Lines et al., 2002; Ramaesh et al., 2003; Smith et al., 2000). However, the severity of ASD presentation for a given mutation is highly dependent on genetic background (Chang et al., 2001; Mao et al., 2015; Smith et al., 2000). These combined observations therefore demonstrate that ASD presents as a complex spectrum of phenotypes and that modifier genes influence the severity of disease presentation. Furthermore, it is clear that additional genes underlying several ASD syndromes remain unidentified. Identification and characterisation of these unknown disease-causing genes using animal models will therefore facilitate an integrated understanding of the pathogenic mechanisms involved in ASD. Moreover, it is entirely possible that these unknown ASD-causing genes could be the underlying genetic basis for other systemic disorders that present...
Tuberous sclerosis complex (TSC) is a systemic syndrome that is caused by inactivating point mutations in either hamartin (TSC1) or tuberin (TSC2), which leads to sustained activation of the mTORC1 signalling pathway and consequent formation of tumour-like lesions (referred to as hamartomas) in affected organs (Han and Sahin, 2011; Napolioli et al., 2009). TSC patients exhibit complex neurological deficits and half of all affected individuals present with visual system involvement that is best characterised by the presence of hamartomas within the posterior segment of the eye (Mennel et al., 2007; Samueli et al., 2015). However, in isolated cases, some TSC patients also present with anterior segment deficiencies, which suggests that this systemic syndrome also involves ASD in individual cases (Eagle et al., 2000; Gutman et al., 1982; Kranias and Romano, 1977; Lucchese and Goldberg, 1981; Milea and Burillon, 1997; Welge-Lussen and Latta, 1976).

The generation and characterisation of an eye-specific TSC mouse model that recapitulated the classical neuropathological hallmarks of this multiorgan syndrome was recently reported (Jones et al., 2015). That study provided the first major insight into the molecular mechanisms underlying eye development during eye development leads to ASD

The morphological appearance of the anterior eye segment in Tsc1 conditional mouse model was first compared to elucidate the consequences of ablation of Tsc1 within the CM (Fig. 2). The anterior segment of mutant mice had consistent morphological deficits that were fully penetrant on the 129/SvCBAC57BL/6 mixed genetic background (n=17 eyes). Iris hypoplasia was evident in frontal views of enucleated Lhx2-Cre: Tsc1f/f eyes at postnatal day (P)15 that resulted in a 2-fold enlargement of the centrally located pupil compared with control animals (Fig. 2A,C and Fig. S2). Moreover, lateral views of the enucleated eyes demonstrated that the anterior eye chamber in Lhx2-Cre: Tsc1f/f mice was reduced in volume (Fig. 2B,D, double arrows), with the cornea being shorter and exhibiting reduced curvature when compared with control littermates (Fig. 2B,D, black lines). However, the posterior eye segment of Lhx2-Cre: Tsc1f/f mice was larger than that of control animals, as previously documented (Fig. 2B,D, white lines) (Jones et al., 2015).

Histological analysis on sagittal eye sections at P18 (Fig. 2E-J) demonstrated that control mice exhibited a CB with well-defined ciliary processes and that the iris appeared to follow the curvature of the lens, with the SP evident at the distal tip (Fig. 2E). In contrast, both the CB and iris of mutant animals appeared hypoplastic with an apparent lack of ciliary processes and iris extension (Fig. 2F). Higher-magnification images confirmed that the CB of control animals was foliated and consisted of an ordered arrangement of both pigmented (PCE) and non-pigmented ciliary epithelia (NCE). In comparison, the overall CB structure in Lhx2-Cre: Tsc1f/f mice lacked any sign of well-defined ciliary processes with the PCE and NCE appearing to coalesce (Fig. 2H). Finally, higher-magnification images also revealed the SP of control animals as an elongated oval structure located in the distal iris tip (Fig. 2I). In contrast, the iris in Lhx2-Cre: Tsc1f/f mice exhibited a thickened club-like appearance with atrophic SP (Fig. 2J).

Immunohistochemical analyses were performed at P14 to elucidate the underlying molecular aetiology for the hypotrophic appearance of the CB and iris in Lhx2-Cre: Tsc1f/f mice (Fig. 3). These experiments were conducted using antibody markers to demarcate specific structures within the anterior eye segment in mice homozygous for the Tyr' mutation (referred to as Lhx2-Cre: Tsc1f/f+; see Materials and Methods) to avoid the masking of antibody fluorescence in pigmented cells. Overview images showed the presence of Pax6+ cells in the anterior segment of both controlPax6- and Lhx2-Cre: Tsc1f/f+ mice (Fig. 3A,B). Higher-magnification analyses demonstrated that the CB of control animals was well defined, with Pax6+ CE cells organised as foliated ciliary processes (Fig. 3C). However, mutant animals exhibited a disorganised Pax6+ CE that resulted in indistinct ciliary processes.
(Fig. 3D). The IPE of control mice exhibited an ordered layer of Pax6+ cells but the equivalent epithelial layer in Lhx2-Cre:Tsc1f/f mice appeared irregular and disorganised (Fig. 3E,F). Additionally, although the presence of αSMA was detected in the iridal muscles (Fig. 3E-H), the levels of αSMA within Lhx2-Cre:Tsc1f/f mice was noticeably reduced, with the SP in particular being atrophic (Fig. 3G,H). To independently validate the observed SP atrophy, the expression of a mature SP-specific marker caveolin-3 (Cav3) was characterised (Kogo et al., 2006). Expression of Cav3 could not be detected within the SP of Lhx2-Cre:Tsc1f/f mice (Fig. 3I,J).

Taken together, these results demonstrate that conditional deletion of Tsc1 within the CM during eye development leads to CB and iris hypoplasia in postnatal mice. It should be noted that in Lhx2-Cre:Tsc1f/f mice (129/SvCBAC57BL/6 mixed genetic background, n=17 eyes), the CB and iris phenotype was fully penetrant and reproducibly severe in mutant animals. However, in the Lhx2-Cre:Tsc1f/f mice (129/SvCBAC57BL/6NMRI mixed genetic background, n=9 eyes) the severity of the anterior segment phenotype was variable among affected animals, albeit with full penetrance (Fig. S3). For example, although the CB in Lhx2-Cre: Tsc1f/f mice was always hypotrophic, the ciliary processes in some eyes were underdeveloped (Fig. S3F), whereas other eyes had ciliary processes that were barely detectable (Fig. S3R). Moreover, in some individuals, the iris appeared as a shortened structure completely lacking a DP (Fig. S3E,G) whereas other animals exhibited somewhat normal iris extension, albeit with obvious DP atrophy (Fig. S3M,O). Finally, the size and position of the SP was also variable in Lhx2-Cre:Tsc1f/f mice (Fig. S3H,L,P,T). It therefore appears that genetic background influences the presentation of the ASD phenotype observed in this study.

**Conditional deletion of Tsc1 leads to elevation of mTORC1 signalling in tissues underlying ciliary body and iris development**

Elevations in mTORC1 signalling within the CM following Tsc1 ablation could cause the CB and iris phenotype observed in postnatal Lhx2-Cre:Tsc1f/f mice (Fig. S4A). Immunoblot analyses were therefore performed on eye homogenates at E14.5 to confirm conditional deletion of Tsc1. A significant reduction in the amount of hamartin (Tsc1) was detected in Lhx2-Cre:Tsc1f/f mice compared with control animals thus confirming eye-specific ablation of Tsc1 (Fig. S4B,C). The level of phosphorylation of the S6 ribosomal protein Rps6 (pS6) was next assessed since this is an established marker for assessing upregulation of mTORC1 signalling following Tsc1 ablation (Fig. S4A) (Biever et al., 2015; Kwiatkowski et al., 2002; Meikle et al., 2007). Significant increases in the levels of both pS6S235/236 and pS6S240/244 were observed in the eye homogenates of Lhx2-Cre:Tsc1f/f mice, thus confirming upregulation of mTORC1 signalling following Tsc1 deletion (Fig. S4B,C).

Development of the CB and iris in the mouse begins at E12.5 and involves interplay between cells residing in the CM, the overlying POM and LE (Cvekl and Tamm, 2004). It was reasoned that the
observed elevation of mTORC1 signalling (Fig. S4B,C) in one or combinations of these tissues underlay the anterior segment phenotype observed in Lhx2-Cre:Tsc1f/f mice. The distribution of pS6 was therefore assessed within these domains during embryogenesis by immunohistochemistry (Fig. 4A-P). No discernible differences were observed in the levels and distribution of pS6 in the optic cup, the overlying POM or the LE at E10.5 (Fig. 4A,E,I,M). By contrast, a premature increase in both pS6S235/236 and pS6S240/244 was detected within the CM, POM and LE of Lhx2-Cre:Tsc1f/f mice at E12.5 (Fig. 4B,F,J,N). At later embryonic stages, the levels of pS6 S235/236 and pS6 S240/244 continued to be elevated in the CM of mutant mice, whereas levels within the POM and LE appeared to be similar to that observed in control littermates (Fig. 4C,D,G,H,K,L,O,P).

Lineage-tracing experiments demonstrated that Lhx2-Cre transgene expression and consequent Tsc1 ablation was solely restricted to the CM (Fig. 1B). The premature elevation of S6 phosphorylation within the CM of Lhx2-Cre:Tsc1f/f mice at E12.5 was therefore to be expected. However, as pS6 levels were also increased within the POM and LE of mutant animals, this suggests that aberrant paracrine signal(s) originating from the CM initiate a premature increase in mTORC1 signalling within these neighbouring domains (Fig. 4F,N). One pathway that can act in this paracrine manner during anterior segment development is retinoic acid (RA) signalling (Duester, 2009). The distribution of retinaldehyde dehydrogenase (Raldh) transcripts was therefore determined to demarcate areas of active RA synthesis within the developing eye (Cvekl and Wang, 2009; Duuster, 2009; Matt et al., 2005; Molotkov et al., 2006). No discernible differences in the level and distribution of either Raldh3 or Raldh1 transcripts were observed within control and Lhx2-Cre:Tsc1f/f mice at E12.5 (Fig. S5A,B,D,E). Raldh3 transcripts were enriched in the ventral retina but also within the dorsal CM and PC (Fig. S5A,D) whereas Raldh1 was highly expressed in the dorsal retina, the ventral CM and the LE (Fig. S5B,E) (Matt et al., 2005). Moreover, there was also no difference in Raldh1 protein expression within the dorsal retina of control and Lhx2-Cre:Tsc1f/f mice (Fig. S5C,F).

Taken together, these combined analyses demonstrate that conditional deletion of Tsc1 during eye development leads to a reduction in hamartin levels and a subsequent premature increase in mTORC1 activity, in turn, induces mTORC1 signalling within the neighbouring POM and LE. However, non-autonomous phosphorylation of S6 within the POM and LE of Lhx2-Cre:Tsc1f/f mice does not appear to be mediated by paracrine RA signalling originating from the CM.

**Conditional deletion of Tsc1 alters transcriptional programs within the ciliary margin**

Aberrant mTORC1 signalling disturbs molecular homeostasis and this underlies a variety of pathological conditions (Duvel et al., 2010). Alterations in the level and/or pattern of gene expression within the CM, POM and/or LE upon premature mTORC1 activation could underlie the CB and iris phenotype observed in...
A comparable spatial distribution of Otx2 transcripts (Martinez-Morales et al., 2001) was observed within the CM of both control and mutant animals at E14.5 and E16.5 (Fig. 6A,B,D,E). However, a reduction in the number of cells expressing the presumptive CM marker msh homeobox 1 (Msx1) (Monaghan et al., 1991) was detected in Lhx2-Cre:Tsc1f/f mice at all ages analysed (Fig. 6G-L). Bone morphogenetic protein (BMP) signalling is required for CB morphogenesis in part by modulation of Msx1 expression (Chang et al., 2001; Zhao et al., 2002). Accordingly, a reduction in the number of cells that express Bmp4 was also observed at E14.5 within the CM of mutant animals when compared with littermate controls (Fig. 6M,P), but no discernible difference in the number of Bmp4-expressing cells was seen at E16.5 (Fig. 6N,Q). Moreover, it was also confirmed that CM length, as defined by the transcriptional domains for all genes, was always consistently shorter in Lhx2-Cre:Tsc1f/f mice at E18.5 (Fig. 6C,F,I,L,O,R and Fig. S6, brackets).

Premature mTORC1 signalling was also observed within the POM and LE of Lhx2-Cre:Tsc1f/f mice (Fig. 4F,N). We therefore assessed whether aberrant gene networks within these domains also contributed to the CB and iris phenotype observed in mutant animals. Particular focus was given to genes that are predominantly expressed for any of these genes (Figs S7 and S8). The expression patterns of Pitx2, Foxc1 and Wnt2b in addition to the Wnt signalling pathway inhibitor 2) and Axin2 were therefore analysed. No differences in transcript distribution within the POM or LE were observed for any of these genes (Figs S7 and S8).

Taken together, these data demonstrate that conditional deletion of Tsc1 during eye development leads to a reduction in the proportion of Pax6+ cells in the CM. This, in combination with a decrease in the number of cells that express Bmp4 and Msx1, presumably leads to the consistent reduction in overall CM length observed at E18.5. Moreover, it appears that CM cell-autonomous mechanisms predominantly underlie the CB and iris phenotype observed in postnatal Lhx2-Cre:Tsc1f/f mice since no changes in the transcriptional distribution of selected candidate genes was observed within the POM and LE.

Conditional deletion of Tsc1 disrupts postnatal ciliary body and iris development

Previous studies have demonstrated that decreases in Pax6 levels reduce the proliferation of CM cells (Davis et al., 2009; Davis-Silberman et al., 2005). Since a reduced number of Pax6+ cells was observed within the CM of Lhx2-Cre:Tsc1f/f mice (Fig. 5N), it was
reasoned that one contributing factor to the failure of CB and iris morphogenesis in postnatal animals was a reduced rate of proliferation within the CM. Late stage embryos (E18.5) were therefore labelled with BrdU and label incorporation was determined at P0. The number of nuclei within the CM of control Tyrc and Lhx2-Cre:Tsc1f/f:Tyrc mice was first quantified and mutant animals were found to contain 24% fewer DAPI+ cells within this domain (Fig. 7A,C,D). However, the CM of mutant animals also demonstrated a 72% reduction in the number of BrdU+ cells, thus demonstrating a reduced rate of progenitor cell proliferation upon Tsc1 ablation (Fig. 7B,E,F).

CB and iris morphogenesis occurs during the first postnatal week of mouse development (Napier and Kidson, 2007). Given that a decrease in overall CM length and progenitor proliferation was observed in Lhx2-Cre:Tsc1f/f mice it was next assessed how this reduction influenced CB and iris development. Histological analysis of sagittal eye sections taken from control mice demonstrated that the future CB was initially demarcated by the emergence of the ciliary processes at P0 (Fig. 7G) and by the completion of the first postnatal week they appeared as distinct foliated structures (Fig. 7I). Simultaneously, the iris elongated toward the centre of the lens and the SP became apparent (Fig. 7I). Thus, by P7, the overall structure of both the CB and iris were well established in control animals (Fig. 7G-I). In contrast, the CB and iris in Lhx2-Cre:Tsc1f/f mice failed to undergo morphogenesis, as illustrated by the absence of ciliary processes coupled with a lack of iris extension (Fig. 7J,K). Failure to reach these developmental milestones at P0 and P3 eventually culminated in the hypotrophic appearance of both the CB and iris at P7 (Fig. 7L).

Taken together, these data demonstrate that conditional deletion of Tsc1 during eye development leads to a reduced number of progenitor cells within the CM of Lhx2-Cre:Tsc1f/f mice, with the remaining cells exhibiting a decreased rate of proliferation. This reduced proliferation rate subsequently leads to a failure of CB and iris morphogenesis during the first postnatal week in mutant animals and eventually culminates in the ASD phenotype observed in Lhx2-Cre:Tsc1f/f mice.

DISCUSSION
This study characterises a novel mouse model of ASD. Conditional deletion of Tsc1 during eye development leads to a premature upregulation of mTORC1 activity within tissue domains whose interplay underlies anterior segment morphogenesis. This aberrant mTORC1 signalling leads to cell-autonomous alterations in genetic networks within the CM and a decrease in cell proliferation with consequent CB and iris hypoplasia in postnatal animals (Fig. 8A,B). Further characterisation of the Lhx2-Cre:Tsc1f/f mouse is required in order to precisely define which particular ASD syndrome the model best represents. These studies could encompass histological and molecular analyses of the lens, cornea and associated drainage structures since aberrant development of these anterior eye tissues...
also contributes to ASD presentation (Idrees et al., 2006; Ito and Walter, 2014). But the neuroectodermal origin of the affected genetic networks within the CM and consequent anterior segment phenotype observed in \textit{Lhx2-Cre:Tsc1f/f} mice is strikingly similar to the underlying molecular aetiology and clinical presentation documented for individuals with aniridia (Beauchamp and Meisler, 1986; Lee et al., 2008). Moreover, as some TSC patients present with anterior segment deficiencies such as aniridia, this suggests that one facet of this systemic syndrome involves ASD in some affected individuals (Eagle et al., 2000; Gutman et al., 1982; Kranias and Romano, 1977; Lucchese and Goldberg, 1981; Milea and Burillon, 1997; Welge-Lussen and Latta, 1976). Given these corroborative experimental and clinical observations, we therefore propose that the \textit{Lhx2-Cre:Tsc1f/f} mice described in this study should be added to the growing list of established ASD models (Fig. 8C) (Gould and John, 2002).

\[\text{Pax6 is a master regulator of eye morphogenesis (Gehring and Ikeo, 1999). This is best exemplified by point mutations in the human } \textit{PAX6} \text{ gene leading to aniridia (Lee et al., 2008), while modulation of mouse Pax6 levels reduces cell proliferation and leads to CB and iris hypoplasia (Davis et al., 2009; Davis-Silberman et al., 2005; Lee et al., 2008; Marquardt et al., 2001). The reduced number of proliferative Pax6+ cells residing in the CM during embryogenesis is therefore presumably the main driving mechanism underlying the anterior segment phenotype observed in postnatal \textit{Lhx2-Cre:Tsc1f/f} mice. However, the precise molecular} \]
Fig. 6. Conditional deletion of Tsc1 disrupts transcriptional programs within the ciliary margin. (A-F) In situ hybridisation analysis of coronal eye sections taken from control (A-C) and Lhx2-Cre:Tsc1f/f (D-F) mice at E14.5 (A,D), E16.5 (B,E) and E18.5 (C,F) demonstrates a comparable expression of Otx1 within the CM at E14.5 (A,D) and E16.5 (B,E). Also note the expression of Otx1 in the overlying PC at E14.5 (A,D). (G-L) In situ hybridisation analysis of coronal eye sections taken from control (G-I) and Lhx2-Cre:Tsc1f/f (J-L) mice at E14.5 (G,J), E16.5 (H,K) and E18.5 (I,L) demonstrates a reduction in the number of Msx1-expressing cells within the CM of mutant animals at all ages analysed. (M-R) In situ hybridisation analysis of coronal eye sections taken from control (M-O) and Lhx2-Cre:Tsc1f/f (P-R) mice at E14.5 (M,P), E16.5 (N,O) and E18.5 (O,R) shows a reduction in the number of cells that express Bmp4 within the CM of mutant animals at E14.5 (M,P). No difference in Bmp4 expression is seen at E16.5 (N,O). In addition, a reduction in total CM length at E18.5 is consistently observed in Lhx2-Cre:Tsc1f/f mice when compared with control animals. CM length is defined as beginning from the distal tip of the CM to the proximal part of the Otx1, Msx1 and Bmp4 expression domains (C,F,I,L,O,R, brackets). Scale bar: 50 µm (A-R).
mechanism by which aberrant mTORC1 activity induces this reduction in CM progenitor cell number remains to be determined. But these findings do concur with recent reports where conditional deletion of Tsc1 or Tsc2 results in a decrease in the number of progenitor cells and consequent tissue hypoplasia during mammary gland and lung development (Qin et al., 2016; Ren et al., 2016). It is also interesting to note that contrasting segment-specific phenotypes exist within Lhx2-Cre:Tsc1f/f mice. A previous report demonstrated that neuroectodermal Tsc1 ablation leads to retinal hyperplasia and corroborates the effects of sustained mTORC1 signalling in other TSC-affected organs (Jones et al., 2015) whereas we show here that conditional deletion of Tsc1 within the same progenitor pool leads to hypoplasia within the anterior segment. Why contrasting compartment-specific developmental deficits exist within the eye of Lhx2-Cre:Tsc1f/f mice is therefore an intriguing avenue for future investigation. However, the CB hypoplasia seen in Lhx2-Cre:Tsc1f/f mice cannot merely be attributed to a reduction in the number of Pax6+ cells alone: other transcription factors and/or

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**Fig. 7. Conditional deletion of Tsc1 disrupts postnatal ciliary body and iris development.** (A-F) CM proliferation rate analysis on coronal eye sections taken from controlTyrc (A,B) and Lhx2-Cre:Tsc1f/f (D,E) mice at P0. Immunohistochemical analysis for neuronal β-III-tubulin was used to define the NR and an ROI was drawn around the presumptive CM (A,D, dashed line). The number of DAPI+ cells was then quantified within this ROI. Lhx2-Cre:Tsc1f/f mice (cKO) had a significant reduction in the total number of DAPI+ cells within the CM at P0 when compared with control littermates (C). Immunohistochemical analysis for BrdU was performed on adjacent sections and the previously drawn ROI was used to define the CM (B,E, dashed line). The number of BrdU+ cells was then quantified within this ROI. This data was subsequently divided by the DAPI+ counts to determine the percentage of BrdU+ cells within the ROI. Lhx2-Cre:Tsc1f/f mice (cKO) had a significant reduction in the percentage of proliferating cells in the CM at P0 when compared with controlTyrc littermates (F). All quantification data are mean±s.e.m. (n=6 eyes for both controlTyrc and Lhx2-Cre:Tsc1f/fTyrc); ****P≤0.0001, calculated using an unpaired two-tailed Student’s t-test. (G-L) Histological analysis of coronal eye sections taken from control (G-I) and Lhx2-Cre:Tsc1f/f (J-L) mice at P0 (G,J), P3 (H,K) and P7 (I,L). The CB and iris undergo gradual morphogenesis in control animals during the first postnatal week (G-I). The future CB is initially demarcated by the emergence of the ciliary processes at P0 (G). Further development of the ciliary processes coupled with IS and IPE extension in addition to the appearance of the SP is observed at P3 (H) and by P7, the overall structure of both the CB and iris are well defined (I). In contrast, the CB and iris fail to undergo morphogenesis during the first postnatal week in Lhx2-Cre:Tsc1f/f mice (J-L). This is best illustrated by the absence of well-defined ciliary processes at P0 (J) coupled with a failure of iris extension and SP development at P3 (K). This failure of morphogenesis eventually culminates in the hypotrophic appearance of both the CB and iris at P7 (L). Scale bars: 25 µm (A,B,D,E), 100 µm (G,H,J,K) and 200 µm (I,L).
signalling molecules probably contribute to this particular phenotype. Possible candidates that fulfil this role are Bmp4 and Msx1 since both of these genes are indispensable for anterior segment morphogenesis (Chang et al., 2001; Mao et al., 2015).

A reduced number of cells expressing Bmp4 and Msx1 was detected in the CM of Lhx2-Cre:Tsc1f/f mice therefore presumably contributes to the CB phenotype. However, it remains to be determined whether the observed CB hypoplasia involves interplay between Pax6, Bmp4 and Msx1 or whether these factors mediate separate developmental deficits.

Identification and characterisation of these modifier genes in different genetic backgrounds may therefore provide an important route for understanding the cellular and genetic interactions underlying the variations in ASD presentation observed in Lhx2-Cre:Tsc1f/f mice on the two mixed genetic backgrounds employed in this study. However, we cannot completely eliminate the possibility that unique stochastic developmental effects within individual embryos also contribute to the divergent spectrum of CB and iris phenotypes seen in these mutant mice (Gould and John, 2002). It is entirely possible that variations in the level of active mTORC1 signalling within individual embryos influence the temporal and spatial activation of anterior segment developmental genes. Appropriate activation of these regulators in some cells and incorrect and/or no activation in others may therefore contribute to the phenotypic variability observed in littermates on the 129/SvCBAC57BL/6NMRI mixed genetic background.

Activation of the ROSA26R construct by Lhx2-Cre-mediated recombination precisely defined the contribution of optic cup progenitor cells to specific anterior segment structures. To the best of our knowledge, this lineage-tracing analysis is the first clear genetic demonstration that progenitor cells residing within the CM of the embryonic mouse eye contribute the epithelial layers of both the CB and iris in addition to the iridial muscles. These observations substantiate formative histological analyses of CB and iris development and confirm that species-specific differences exist among anterior segment development (Beebe, 1986; Imaiuzumi and Kuwabara, 1971). This is best exemplified by our results demonstrating that the mouse iridial muscles are derived from optic cup progenitor cells, whereas these structures originate from the neural crest in birds (Creuzet et al., 2005; Johnston et al., 1979; Nakano and Nakamura, 1985). The Lhx2-Cre transgene is therefore a powerful molecular tool that can be used to elucidate the genetic networks that underlie the development and disease of both posterior and anterior segment structures that arise from neuroectodermal progenitor cells. Moreover, combinatorial analyses using the Lhx2-Cre transgene in conjunction with other compatible lineage-tracing reporter systems would be an invaluable approach to gain a better understanding of how cells originating from different embryonic regions simultaneously migrate to assemble the eye (Legue and Joyner, 2010).

To conclude, this study describes the characterisation of a novel mouse model of ASD. This model provides a valuable resource for future studies concerning the molecular mechanisms underlying ocular syndromes and also serves as a platform to evaluate new therapeutic approaches for the treatment of visual disorders. In addition, the combined studies on anterior and posterior segment development in Lhx2-Cre:Tsc1f/f mice (Jones et al., 2015) demonstrate that the various tissues contributing to the formation of the adult mouse eye require separate and distinct levels of mTORC1 signalling for correct morphogenesis.

**MATERIALS AND METHODS**

**Animals**

All animal experiments were approved by the animal review board at the Court of Appeal of Northern Norrland in Umeå. The derivation and genotyping of Tg(Lhx2-Cre)1Lcar transgenic mice (abbreviated to Lhx2-Cre), Tsc1tm1Djk floxed mice (Tsc1f/f or Tsc1f/f) and Gt(Rosa)26Sor1Cre reporter mice (ROSA26R) have been described previously (Hägglund et al., 2011; Soriano, 1999; Uhmann et al., 2002). The genotype of all animals was determined by PCR analysis of genomic DNA extracted from tail biopsies. Breeding Lhx2-Cre:Tsc1f/f and Tsc1f/f mice or Lhx2-Cre and ROSA26R mice generated all experimental animals. The morning of the vaginal plug was considered as E0.5. Both males and females were used for experimental analyses and littermates lacking the Lhx2-Cre transgene were
used as controls. All analyses were carried out on a 129/SvBCAC57BL/6 (control and Lhx2-Cre:Tsc1f/f) and/or 129/SvBCAC57BL/6NMR1 mixed genetic backgrounds. The latter strain was generated by crossing 129/SvBCAC57BL/6 mice with B6Tact:NMRI mice, which lack pigment due to a mutation in the Tyr gene (Tyr<sup>−</sup>). The 129/SvBCAC57BL/6NMR1 mixed background strain was then bred to be homozygous for the Tyr<sup>−</sup> mutation (control<sup>Tyr<sup>−</sup></sup> and Lhx2-Cre:Tsc1f/f<sup>Tyr<sup>−</sup></sup>). All postnatal analyses of Lhx2-Cre:Tsc1f/f mice were conducted before P21 since this model dies from neurological complications at approximately 3 weeks of age (Jones et al., 2015).

**Histology**
Heads or enucleated eyes were fixed in 2% (w/v) glutaraldehyde and 3% (w/v) paraformaldehyde (PFA) in PBS overnight at 4°C. Tissues were then either immersed in 70% (v/v) ethanol and paraffin embedded or equilibrated in 30% (w/v) sucrose in PBS overnight at 4°C and embedded in OCT compound (Sakura Finetek). Paraffin sections (10 µm) were subsequently cleared in xylene (2×5 min) before rehydration through a series of ethanol washes [99.5%, 95%, 90% and 80% (v/v) in PBS]. Cryosections (10 µm) were rehydrated in PBS (2×5 min). Haematoxylin and Eosin staining was performed as previously described (Hägglund et al., 2011).

**Lineage tracing**
Heads or enucleated eyes were fixed in 4% (w/v) PFA in PBS for 30 min on ice and then equilibrated in 30% (w/v) sucrose in PBS overnight at 4°C. Tissues were embedded in OCT compound (Sakura Finetek) and cryosections (10 µm) were prepared. Lineage-tracing analyses were performed as described previously (Hägglund et al., 2011).

**Immunohistochemistry**
Heads or enucleated eyes were fixed in 4% (w/v) PFA in PBS for up to 2 h on ice. The tissues were equilibrated overnight at 4°C in 30% (w/v) sucrose in PBS and embedded in OCT compound (Sakura Finetek). Immunohistochemistry was performed on cryosections (10 µm) as previously described (Ma et al., 2007). An additional blocking step involving MOM Blocking Reagent (Vector Labs) was used in all experiments involving monoclonal primary antibodies. The following antibodies and dilutions were used: aSMA (1:200, Abcam, ab5694 and 1:500, Sigma-Aldrich, C6198), β-gal (1:500, MP Biomedical, 0855976); Cav3 (1:500, BD Biosciences, 610421), neuronal β III-tubulin (1:1000, Covance, MRB-435), Pax6 (1:100, DSHB, Clone PSU1), pS6<sup>T235/236</sup> (1:100, Cell Signaling Technology, 4857), pS6<sup>T240/244</sup> (1:100, Cell Signaling Technology, 1217), Raldh1 (1:4000, Abcam, ab96060). All immunohistochemistry antibodies have been independently verified in a previous study (Jones et al., 2015).

**Immunoblotting**
The lens was removed from enucleated eyes and the NR and RPE frozen in liquid nitrogen. Tissues were homogenised in SDS lysis buffer [100 mM Tris-HCl, pH 6.8; 2% (w/v) SDS] containing both protease and phosphatase inhibitors (Complete Mini & PhosSTOP, Roche) using a TissueLyser (Qiagen). The concentration of the soluble protein fraction was measured using the BCA Protein Assay Kit (Fisher Thermo Scientific). Immunoblotting was performed on protein extracts (6 µg) using Criterion TGX gels and nitrocellulose membranes (Bio-Rad) (Mahlmod and Yang, 2012). Reactive proteins were visualised using SuperSignal West Dura Extended Duration Substrate (Fisher Thermo Scientific). Quantification was performed using a ChemiDoc MP Imaging System (Bio-Rad). The following antibodies and dilutions were used: Hamartin (1:1000, Cell Signaling Technology, 4906), S6 (1:2000, Cell Signaling Technology, 2217), pS6<sup>T235/236</sup> (1:1000, Cell Signaling Technology, 12468), pS6<sup>T240/244</sup> (1:1000, Cell Signaling Technology, 12469) and GAPDH (1:30,000, Cell Signaling Technology, 2118). All immunoblotting antibodies have been verified in a previous study (Jones et al., 2015).

**In situ hybridisation**
Heads were fixed in 4% (w/v) PFA in PBS for 2 h on ice and equilibrated in 30% (w/v) sucrose in PBS overnight at 4°C. Tissues were embedded in OCT compound (Sakura Finetek). **In situ** hybridization on cryosections (10 µm) was performed as previously described (Scharen-Wiemers and Gerfin-Moser, 1993). The following cDNA templates or IMAGE clone were used to generate riboprobes: Axin2 (BC057338, nucleotide position 1-1520), Bmp4 (NM_007554, nucleotide position 117-578), Bmp7 (NM_007557, nucleotide position 1-1987), Dkk2 (BC096448, nucleotide position 1-3713), Foxc1 (NM_008592, nucleotide position 526-1354), Msx1 (NM_010835, nucleotide position 274-1138), Otx1 (NM_011023, nucleotide position 168-962), Pdz2 (NM_001042504, nucleotide position 59-799), Raldh1 (BC054386, nucleotide position 755-2078), Raldh3 (NM_053080, nucleotide position 247-1088), Rhl10 (NM_133832, nucleotide position 14-923), Sfppl (NM_013834, nucleotide position 109-923), Wnt2b (IMAGE clone #40057540) and Zic1 (NM_009573, nucleotide position 272-1176).

**Proliferation analyses**
Pregnant dams were injected intraperitoneally at E18.5 with BrdU at 50 µg/g body weight and pups were collected at P0. Heads were fixed in 4% (w/v) PFA in PBS for 2 h on ice and then equilibrated overnight at 4°C in 30% (w/v) sucrose in PBS before embedding in OCT compound (Sakura Finetek). Cryosections (10 µm) were washed in PBS and then denatured in 2 M HCl for 30 min at 37°C. The slides were subsequently neutralised with 100 mM borate buffer (pH 8.5) for 10 min at RT and then processed for immunohistochemistry as previously described using an anti-BrdU antibody (1:20, BD Pharmingen, 562010) (Jones et al., 2015).

**Image analyses**
Images were captured using either a Zeiss LSM 710 confocal microscope, a Nikon Eclipse E800 microscope fitted with a Nikon DS-Ri1 digital colour camera or a Nikon SMZ1500 microscope fitted with a Nikon DS2 digital colour camera. All images were compiled and analysed using Fiji (Schindelin et al., 2012), CellProfiler (Carpenter et al., 2006; Kamentsky et al., 2011), Adobe Photoshop and Adobe Illustrator.

**Statistical analyses**
All statistical analyses were performed using Prism7 (GraphPad). Quantification analyses were performed blind to genotype and unpaired two-tailed Student’s t-tests were used to determine statistical significances. Error bars in all figures represent s.e.m. P-values are indicated as *P<0.05, **P<0.01, ***P<0.001 and ****P<0.0001.

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**Competing interests**
The authors declare no competing or financial interests.

**Author contributions**
A.-C.H., I.J. and L.C. designed and conceived the experiments. A.-C.H. and I.J. performed the experiments. A.-C.H., I.J. and L.C. analysed the data. A.-C.H., I.J. and L.C. wrote the paper.

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**Supplementary information**
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**References**


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